

Structural Determination of (—)-SCH 64874 and Hirsutellomycin by Semisynthesis

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Supporting Information

ABSTRACT: The structure of a C_2 -symmetric epidithiodike-topiperazine alkaloid, SCH 64874, was determined by semisynthesis. The relative stereochemistry of the β -hydroxy carboxylic acid chain having three chiral centers was determined by comparison of the NMR data of the four possible diastereomeric β -hydroxy carboxylic acid fragments

with those of SCH 64874. Condensation of the (–)-deacetylaranotin core with two enantiomeric β -hydroxy carboxylic acids revealed the relative stereochemistry of SCH 64874. The relative stereochemistry of the β -keto carboxylic acid chain of the analogous alkaloid hirsutellomycin was determined in a stepwise manner. The C4′–C6′ syn relationships were predicted by comparing the NMR data of the corresponding ester fragments with that of hirsutellomycin. The relative stereochemistry of the whole molecule, including the epimerizable C2′ stereocenter, was determined by introduction of four possible side chains into the bisdethiodi(methylthio)deacetylaranotin core. We found that the stereochemistry of C2′ converged with that of the thermodynamically stable form influenced by the core structure.

INTRODUCTION

Epidithiodiketopiperazine (ETP) alkaloids have attracted considerable attention because of their unique structures possessing a sulfur-bridged diketopiperazine core and intriguing biological activities. Among them, seven-membered dihydrooxepine-fused pyrrolidine/ETPs such as (–)-acetylaranotin (1), (–)-emethallicin A (2), and (–)-MPC1001 (3) have been thoroughly investigated for the last few decades, since they exhibit fascinating biological activities (Figure 1). In spite of these attractive features, synthetic difficulties and a scarcity of natural compounds prevented this class of compounds from being further studied even in terms of their structural determination.

(-)-SCH 64874 (4),⁵ isolated from the organic extract of the fermentation broth of an unidentified fungus, is one such compound that has been investigated due to its fascinating biological activities. This compound displays potent epidermal growth factor receptor antagonist activity. Although Hegde and co-workers presumed the structure of 4 to possess a (-)-deacetylaranotin (5) core, the absolute stereochemistry of the molecule and relative stereochemistry of the side chain remain unknown. Hirsutellomycin (6),6 which had been isolated from the submerged cultures of the entomopathogenic fungus Hirsutella kobayasii BCC 1660 by former colleagues of one of the authors of this paper (M.I.) at the BIOTEC, Thailand, was reported to possess potent antituberculosis capabilities. However, only the planar structure was investigated, and further biological activities have not yet been studied; this is due to the exhaustion of the supply of natural

Figure 1. Epidithiodiketopiperazines (ETPs).

samples, and because the strain of *H. kobayashi* which produced **6** ceased to be in production. For accelerating further medical and biochemical investigations on these promising compounds, structural determination based on a synthetic approach is unavoidable. In this paper, we describe the semisynthesis of **4**

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and 6, aiming at the elucidation of the structure of these compounds.

RESULTS AND DISCUSSION

Structural Determination of SCH 64874. We designed a strategy for the semisynthesis of structurally unidentified SCH 64874 (4) so that synthetic efforts would become as efficient as possible (Scheme 1). Considering the C_2 symmetry, 5 4 can be

Scheme 1. Strategy for Structural Determination of SCH 64874 (4)

divided into (-)- or (+)-5 and the two equivalent carboxylic acids 7, which should be one of the eight possible stereoisomers (Scheme 1a). Due to the limited availability of (-)-1, a precursor of (-)-5, by the fermentation and isolation process, we used the following efficient strategy to semisynthesize (-)-4 using a minimum amount of the deacetylaranotin core (-)-5. Thus, we tried to predict the relative stereochemistry of carboxylic acid 7 by comparing the NMR spectra of four possible diastereomeric esters 8a-d with those of the corresponding side chain of the natural compound (Scheme 1b). Then, condensation of (-)-5 with two enantiomers of carboxylic acid 7 possessing the predicted relative stereochemistry should reveal the relative stereochemistry between the aranotin core and the side chain and should reveal the absolute stereochemistry of 4 by comparing the optical rotation with the natural compound.

Esters 8a and 8b were synthesized using the Evans aldol reaction (Scheme 2). Treatment of (-)-9 with $n\text{-Bu}_2\text{BOTf}^8/$ Et₃N, followed by addition of aldehyde (+)-10, afforded aldol product 11a as a single diastereomer. The chiral auxiliary was removed by methanolysis to give ester 8a. Similarly, a combination of the boron enolate prepared from (+)-9 and (+)-10 afforded aldol product 11b, which was converted to ester 8b by methanolysis.

Conversely, esters $8c^{11}$ and 8d were synthesized by a non-Evans anti-type aldol reaction (Scheme 3). Treatment of (+)-9 with n-Bu₂BOTf/i-Pr₂NEt, followed by addition of (+)-10 in the presence of SnCl₄, afforded aldol product 11c as a 4:1 diastereomeric mixture. After the separation of the minor diastereomer, the chiral auxiliary was removed by methanolysis to furnish ester 8c. Ester 8d was also synthesized via aldol product 11d by the same protocol starting from (-)-9.

Scheme 2. Synthesis of Esters 8a and 8ba

^aReagents and conditions: (a) n-Bu₂BOTf, Et₃N, CH₂Cl₂, −78 to 0 °C then (+)-**10**, −78 to 0 °C, 63% (single diastereomer); (b) NaOMe, MeOH, 0 °C, 91%; (c) n-Bu₂BOTf, Et₃N, CH₂Cl₂, −78 to 0 °C; (+)-**10**, −78 to 0 °C, 83% (single diastereomer); (d) NaOMe, MeOH, 0 °C, 81%.

Scheme 3. Synthesis of Esters 8c and 8d^a

^aReagents and conditions: (a) n-Bu₂BOTf, i-Pr₂NEt, CH₂Cl₂, -78 to 0 °C; (+)-10, SnCl₄, -78 to 0 °C, dr = 4:1; separation, 47%; (b) NaOMe, MeOH, 0 °C, 82%; (c) n-Bu₂BOTf, i-Pr₂NEt, CH₂Cl₂, -78 to 0 °C; (+)-10, SnCl₄, -78 to 0 °C, dr = 23:20; separation, 36%; (d) NaOMe, MeOH, 0 °C, 64%.

With the requisite diastereomers 8a-d in hand, a careful NMR analysis was performed to predict the relative stereochemistry of the side chain of the natural product. Initially, differences in ¹³C NMR chemical shifts between the side chains of 4 and each of the diastereomers 8a-d were calculated (Figure 2). Differences between the ¹³C NMR spectra of 4 and those of 8a-c were smaller than those of 8d. Next, we compared their ¹H NMR spectra to that of 4 (Figure 3). It was observed that the ¹H NMR spectrum of 8a was very close to that of 4, whereas that of 8b-d had a big difference regarding the 3' and/or 5' position. These results clearly indicated that the relative stereochemistry of the side chains of 4 corresponds to that of 8a.

Finally, both carboxylic acid (-)-13 and its enantiomer were introduced onto the (-)-deacetylaranotin (5) core to determine the relative and absolute stereochemistry of 4. THP-protected β -hydroxy carboxylic acids (-)-13 and (+)-13 were prepared from 11a and 11e, ¹³ respectively, by THP protection followed by the removal of the chiral auxiliary (Scheme 4).

The (-)-deacetylaranotin (5) core was prepared from (-)-acetylaranotin (1), which was isolated from fermentation broth of *Aspergillus terrreus* BCC 4480. Treatment of 1 with a weak acid (0.9% aq. HCl in MeOH) afforded (-)-5 without the loss of the disulfide bridge (Scheme 5). (-)-Diol 5 was condensed with two carboxylic acids (-)-13 using WSCD·

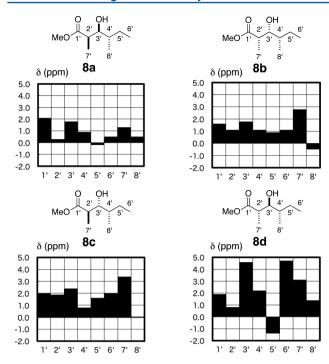


Figure 2. Comparative study of ¹³C NMR data (100 MHz, CDCl₃).

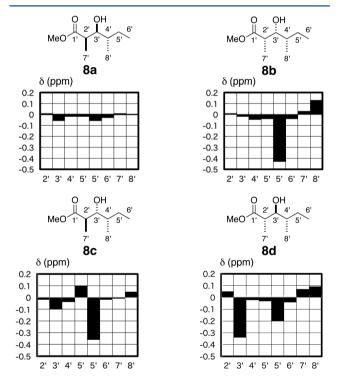


Figure 3. Comparative study of ¹H NMR data (400 MHz, CDCl₃).

HCl/DMAP to give diester 14 in moderate yield. Finally, the THP groups were removed by a careful treatment with PPTS¹⁵ to obtain 15. In addition, 17 was also derived from (-)-5 and two (+)-13 fragments by the same protocol.

The difference between the chemical shifts in the 13 C and 1 H NMR spectra of 4 and synthetic compounds 15 and 17 (Figure 4) clearly demonstrated that the NMR profile of 17 completely matched with 4, indicating that the relative stereochemistry of 4 was that described in 17. In addition, the negative optical rotation of synthetic 17 matched with that of natural 4 ($[\alpha]_{D}^{2p}]$ =

Scheme 4. Synthesis of Carboxylic Acids (-)-13 and (+)-13

^aReagents and conditions: (a) DHP, TsOH·H₂O, CH₂Cl₂, rt, 90%; (b) LiOH·H₂O, H₂O₂, THF-H₂O (7:2), 0 °C to rt, 82%; (c) DHP, TsOH·H₂O, CH₂Cl₂, rt, 76%; (d) LiOH·H₂O, H₂O₂, THF-H₂O (7:2), 0 °C to rt, 84%.

Scheme 5. Synthesis of two Candidates of SCH 64874 (4)^a

"Reagents and conditions: (a) 0.9% aq. HCl in MeOH, rt, 72%; (b) (–)-13 WSCD·HCl, DMAP, CH₂Cl₂, rt, 42%; (c) PPTS, MeOH-CH₂Cl₂, rt, 66%; (d) (+)-13 WSCD·HCl, DMAP, CH₂Cl₂, rt, 38%; (e) PPTS, MeOH-CH₂Cl₃, rt to 40 °C, 86%.

 -169° (c=0.141, CHCl₃), lit. $[\alpha]_{\rm D}^{29}=-301.1^{\circ}$ (solvent and concentration were not reported)),⁵ strongly suggesting that the absolute configuration of 4 should be that of 17.¹⁶

Structural Determination of Hirsutellomycin. The focus of our investigation then moved onto the structural determination of hirsutellomycin (6) by semisynthesis. Due to the unavailability of natural hirsutellomycin (6), our only means of determining its structure was chemical synthesis. Our own analysis of the NMR spectra (¹H NMR, ¹³C NMR, DEPTs, COSY, HMQC, and HMBC) provided by the isolation chemists agreed with the reported planar structure of 6. The close resemblance of the NMR data obtained for natural hirsutellomycin (6) with those of acetylaranotin and SCH 64874 (17) suggested that 6 shares the same relative configuration of it is deacetylaranotin core. Unfortunately, due to a lack of data regarding its optical rotation, it was impossible to determine the absolute stereochemistry of 6.

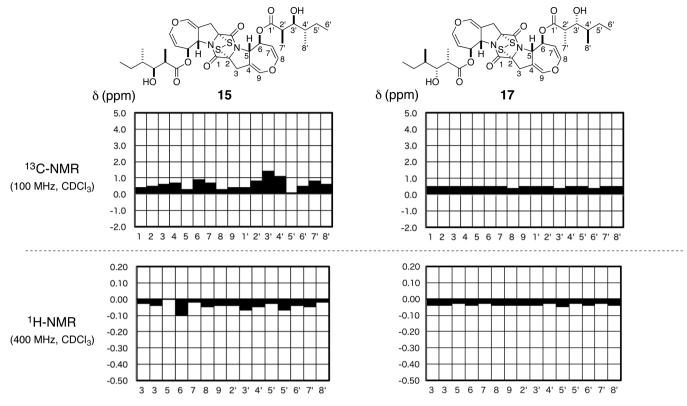
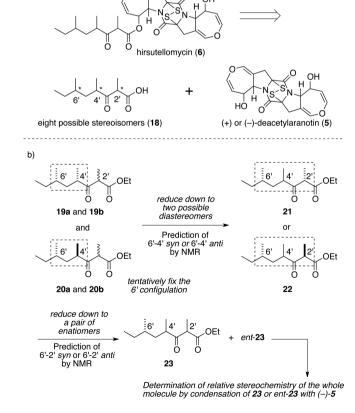


Figure 4. Comparative study of NMR data.

However, the fact that all of the analogous fungal epipolythiodiketopiperazines, including 1, 2, 3, 17, aranotin, and apoaranotin, contain (–)-deacetylaranotin-like (dihydroox-epine-containing) cores, while not containing its antipode, strongly suggests that hirsutellomycin (6) should also have a (–)-deacetylaranotin core. Based on this hypothesis, we planned a semisynthesis strategy similar to the one used for SCH 64874 using a (–)-deacetylaranotin (5).

We planned an efficient strategy for the semisynthesis of the structurally unidentified compound 6 (Scheme 6). A disconnection in the ester moiety of **6** gives a β -keto carboxylic acid fragment 18 (Scheme 6a). In order to avoid the tedious preparation of the eight possible enantiomers of 18, we used Professor Kishi's Universal NMR Data Base concept¹⁷ to try to predict the relative stereochemistry C6'-C4' using simple ester fragments. The concept states that the stereoelectronic interactions and the spectroscopic properties between the structural clusters in a molecule depend on the substituents that are connected either directly to or with a one-methylene bridge that does not depend on the rest of the molecule. 17 Thus, the absolute configuration of C6' was tentatively fixed to be S, and the relative configuration between C6'-C4' was predicted by comparing the NMR data of the ethyl esters 19 and 20 with the corresponding part of the ester chain of the natural compound 6 (Scheme 6b). This would reduce the four possible diastereomers down to just two (21 or 22). The prediction of the relative stereochemistry between C2' and C6' would further reduce the possibility down to just one diastereomer and its enantiomer. Finally, a condensation of the naturally occurring (-)-deacetylaranotin core with 23 and ent-23 revealed the diastereomeric relationship between the side chain and the deacetylaranotin core, respectively, which could be used to determine the relative stereochemistry of 6.

Scheme 6. Strategy for Structural Determination of Hirsutellomycin (6)



Esters 19 and 20 were prepared as diastereomeric mixtures regarding the C2′position from the known compounds of 24¹⁸ and 28,¹⁹ respectively (Scheme 7). Removal of the

Scheme 7. Synthesis of β -Keto Esters 19 and 20.^a

"Reagents and conditions: (a) LiBH₄, THF-MeOH, 0 °C to rt; (b) SO₃-Py, Et₃N, CH₂Cl₂-DMSO, 0 °C; (c) **27**, SnCl₂, CH₂Cl₂, rt, 24% (3 steps, **19a**:19b = 1:1); (d) NaHMDS, THF,—78 °C; MeI, 71% (single diastereomer); (e) LiBH₄, THF-MeOH, 0 °C to rt; (f) SO₃-Py, Et₃N, CH₂Cl₂-DMSO, 0 °C to rt; (g) **27**, SnCl₂, CH₂Cl₂, rt, 20% (3 steps, **20a**:20b = 1:1).

oxazolidinone with LiBH₄ provided alcohol **25**, which was then subjected to Parikh—Doering oxidation²⁰ to give aldehyde **26**. Treatment of aldehyde **26** with diazoester **27** afforded the desired 4′,6′-syn ethyl ester **19a** and **19b** as a 1:1 diasteromiric mixture.²¹ On the other hand, diastereoselective methylation of **28** gave **29** as a single diastereomer. As described in the synthesis of **19**, **29** was converted to a 1:1 diastereomeric mixture of 4′,6′-anti ethyl ester **20a** and **20b**.

A comparison between the ¹H NMR spectra of **6** with those of **19** and **20** suggested that the relative configurations between C-4' and C-6' of the natural product would be *syn* (Figure 5).²² As such, a signal corresponding to the C-5' position of **6** (1.7–1.8 ppm) was also observed for **19** (1.6–1.8 ppm). In contrast,

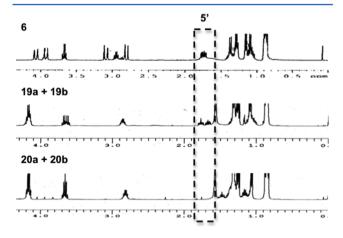


Figure 5. Comparative study of ¹H NMR spectrum (400 MHz, CDCl₃).

the signals on the C-5' of **20** appeared far from those of **19** and **6** (1.48 and 1.25 ppm, respectively).

Next, we attempted to predict the relative stereochemistry between the C4',6', and C2' positions by the stereoselective synthesis of esters 19a (C2'-C6' syn) and 19b (C2'-C6' anti) and the comparison of their NMR data. However, as the stereochemistry of the C2' position was easily epimerized and giving a 1:1 mixture of the diastereomers, we abandoned this strategy (Scheme 8). An attempt was made to prepare 19a by

Scheme 8. Trial for the Synthesis of β -Keto Ester 19a^a

"Reagents and conditions: (a) 31, -78 to 0 °C; separation, 65% (3 steps from 24); (b) LiOH·H₂O, H₂O₂, THF, 0 °C to rt, 86%; (c) K₂CO₃, EtI, acetone, 50 °C, 89%; (d) Dess–Martin periodinane, CH₃Cl₃, 0 °C, 90% (19a:19b = 1:1).

first using the aldol reaction of the aldehyde 26 with a boron enolate 31 to give the aldol 32²³ as the sole product. After the removal of the chiral auxiliary through basic hydrolysis, the resultant carboxylic acid was converted to an ethyl ester 34 under alkylation conditions. Unfortunately, the oxidation of the hydroxyl group with a Dess–Martin periodinane²⁴ gave a 1:1 mixture of 19a and 19b.

The revised strategy is depicted in Scheme 9. We considered that the stereochemistry on the side chain of C2' was strongly

Scheme 9. Strategy for Determination of Relative Configuration

affected by the core skeleton, and so a thermodynamically more stable compound should exist. With this hypothesis, we planned to synthesize a series of compounds 37–40 by introduction of four possible diastereoisomers of side chains on the core skeleton and examine their thermodynamic stability. We chose the more abundant and easily available bisdethiodi(methylthio) deacetylaranotin 35²⁵ (from 36) instead of the

(–)-deacetylaranotin (5) as the core skeleton for this synthesis. In addition to elucidating the relationship between C4′,6′, and C2′, we hoped to obtain information on the relative stereochemistry between the core skeleton and the side chain.

Scheme 10 illustrates one example of introduction of side chain fragments onto the core skeleton 35. The aldol product

Scheme 10. Introduction of the Side Chain Fragments^a

"Reagents and conditions: (a) DHP, PPTS, CH₂Cl₂, rt; (b) LiOH·H₂O, H₂O₂, THF-H₂O, 0 °C to rt, 89% (2 steps); (c) K₂CO₃, MeOH, rt, quant.; (d) **41**, WSCD·HCl, DMAP, CH₂Cl₂, reflux, 36%; (e) PPTS, MeOH-CH₂Cl₂, 40 °C, 78%.

32 was converted into a carboxylic acid 41 via protection of hydroxyl group and hydrolysis of chiral auxiliary. Treatment of diacetate 36 with K_2CO_3 in MeOH produced alcohol 35, which was condensed with carboxylic acid 41 to give the ester 42 in moderate yield. Acidic deprotection of THP group ¹⁵ gave β -hydroxy ester 43.

Other carboxylic acids that were required to synthesize 38, 39, and 40 were synthesized by standard transformations. For example, the preparation of 50 started with the diastereoselective methylation of 44²⁶ to give 45,²⁷ which was subsequently converted to the aldehyde 47 through the removal of the chiral auxiliary and a Parikh–Doering oxidation²⁰ (Scheme 11). After the stereoselective aldol reaction of the aldehyde 47 with a boron enolate 48, the resulting aldol 49 was protected by a THP group,¹⁵ and the oxazolidinone was removed so as to furnish the carboxylic acid 50.

Having prepared these carboxylic acids, these compounds were condensed with the core skeleton 35 through the same protocol used to obtain ester 43 (vide supra). The configurational stability of the C2′ position of the resultant esters was then carefully inspected (Scheme 12).

In all of the cases, the C2' stereochemistry was epimerized so as to preferentially provide the C2'(R) isomer during the Dess–Martin oxidation. Although esters **38** and **40** were partially epimerized, the C2'(R) configuration of esters **37** and **39** tended to easily epimerize the C2'(S) isomers of **38** and **40**. These results indicated that the compounds **38** and **40** were more thermodynamically stable than **37** and **39**, respectively. Consequently, we predicted that the relative configurations of the C2' and C5 positions of hirsutellomycin (**6**) are (R,S) or (S,R), respectively.

Scheme 11. Synthesis of Carboxylic Acid 50^a

"Reagents and conditions: (a) NaHMDS, THF, -78 °C; MeI, 85% (single diastereomer); (b) LiBH₄, THF-MeOH, 0 °C to rt; (c) SO₃· Py, Et₃N, CH₂Cl₂-DMSO, 0 °C; (d) **48**, -78 to 0 °C, 45% (3 steps); (e) DHP, PPTS, CH₂Cl₂, rt; (f) LiOH·H₂O, H₂O₂, THF-H₂O, 0 °C to rt, 65% (2 steps).

Scheme 12. Thermal Stability of 2' Configuration^a

^aReagents and conditions: (a) DMP, CH₂Cl₂, 0 °C, 34% (from 43, 37:38 = 2:3); (b) DMP, CH₂Cl₂, 0 °C, 67% (from 51, 37:38 = 1:5); (c) DMP, CH₂Cl₂, 0 °C, 48% (from 52, 39:40 = 3:2); (d) DMP, CH₂Cl₂, 0 °C, 37% (from 53, 39:40 = 1:5).

The relative configuration between the C4′,6′, and C2′ positions on the side chain was also predicted by comparing the ¹³C NMR spectra of 6 and the model compounds 38 and 40 (Figure 6). By judging the degree of deviation between the model compounds and 6, we predicted that the relative configuration of the side chain of 6 might be 38.

At this stage, we were ready to determine the relative configuration of hirsutellomycin (6) through the semisynthesis

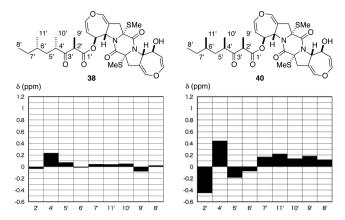


Figure 6. Comparative study of ¹³C NMR data (150 MHz, CDCl₃).

of the predicted structure 56. The synthesis of 57 commenced with the condensation of (–)-5 with the carboxylic acid 54 to provide ester 55 in a 21% yield (Scheme 13). A careful

Scheme 13. Synthesis of β -Keto Ester 57^a

^aReagents and conditions: (a) WSCD·HCl, DMAP, CH₂Cl₂, reflux, 21%; (b) PPTS, MeOH-CH₂Cl₂, 50 °C; (c) DMP, CH₂Cl₂, 0 °C, 58% (3 steps, dr 2′*R*:2′*S* = 10:1; this reaction was carried out once again by using recovered SM).

deprotection of the THP group using PPTS¹⁵ produced the β -hydroxy ester **56**. Finally, the Dess–Martin oxidation of the hydroxyl group in **55** gave the β -keto ester **57** as a 10:1 diastereomeric mixture. The ¹H- and ¹³C NMR spectra of the synthesized compound **57** were completely identical with those of the natural compound **6**.⁶ From this result, therefore, we believe the relative configuration of hirsutellomycin (**6**) to be that of **57**.

CONCLUSION

In conclusion, we accomplished to semisynthesize (–)-SCH 64874 (4) and hirsutellomycin (6) from (–)-deacetylaranotin (5). Based on these semisyntheses, we were able to determine the relative configurations of 4 and 6. During the structural determination of 6, we found that the configuration of the active methine moiety in the side chain of 6 depended on that of the core skeleton. These results are expected to be useful for both synthetic and biological studies of these compounds and their related derivatives as well as compounds that possess a β -keto ester functionality.

■ EXPERIMENTAL SECTION

General Remarks. Materials were obtained from commercial suppliers and used without further purification unless otherwise mentioned. Anhydrous MeOH, Et₃N, and DIPEA were dried and distilled according to the standard protocols. Column chromatography was performed on silica gel 60N (spherical neutral, 63-210 μ m), and flash column chromatography was performed on silica gel 60N (spherical neutral, $40-50 \mu m$). Preparative TLC and analytical TLC were performed on glass plates precoated with a 0.25 mm thickness of silica gel. All melting points were determined on a melting point apparatus and are uncorrected. NMR spectra were recorded on a 400, 500, and 600 MHz spectrometer. Chemical shifts for ¹H NMR are reported in parts per million (ppm) downfield from tetramethylsilane as the internal standard and coupling constants are in Hertz (Hz). The following abbreviations are used for spin multiplicity: s = singlet, d = doublet, t = triplet, q = quartet, m = multiplet, and br = broad. Chemical shifts for 13C NMR are reported in ppm, relative to the central line of a triplet at 77.0 ppm for deuteriochloroform. IR spectra were measured on a FTIR spectrometer. Mass spectra were recorded on a EI or ESI-TOF MS spectrometer. Optical rotations were measured on a polarimeter. The structures of compounds S1-S9 are indicated in the Supporting Information.

Search for Acetylaranotin-Producing Fungal Strains. To find acetylaranotin-producing fungal strains, 10 strains of *Aspergillus terreus*, deposited in the BIOTEC Culture Collection (BCC), were fermented in small scale (250 mL), and their extracts were analyzed by ¹H NMR (CDCl₃). Four strains produced acetylaranotion (1) and bisdethiodi(methylthio) acetylaranotin (36). In all cases, the efficacy of the production of 1 was poor. Although not quantitative, the results suggested that strain BCC 4480 was the most suitable acetylaranotin producer. This fungus was isolated from a soil sample collected in Thailand and identified by Prof. Leka Manoch, and it was deposited in the BCC on August 27, 1998.

Fermentation of Aspergillus terreus BCC 4480 and Isolation of 1 and 36. The fungus BCC 4480 was fermented in Czapek-Dox broth (CZB; sucrose 30.0 g/L, NaNO₃ 3.0 g/L, K₂HPO₄ 1.0 g/L, $MgSO_4.7H_2O$ 0.5 g/L, KCl 0.5 g/L, FeSO₄·7H₂O 0.1 g/L; 200 × 250 mL) at 25 °C for 35 days under static conditions. The cultures were filtered to separate broth (filtrate) and mycelia (residue). The broth (ca. 50 L) was extracted with EtOAc (3 × 50 L) to obtain a brown gum (10.4 g, broth extract). The wet mycelia were macerated in MeOH (5 L, rt, 3 days) and filtered. H₂O (300 mL) and hexanes (1.5 L) were added to the filtrate, and the layers were separated. The aqueous MeOH (bottom) layer was partially concentrated by evaporation, and the residue was extracted with EtOAc. The combined EtOAc solution was concentrated under reduced pressure to obtain a brown gum (1.37 g, mycelial extract). The broth extract was fractionated by column chromatography on Sephadex LH-20 (MeOH), and the fractions containing target compounds were further separated by silica gel column chromatography (MeOH-CH2Cl2) and preparative HPLC using a reversed-phase column (MeOH/H₂O = 45:55) to furnish 1 (10 mg) and 36 (56 mg). The mycelial extract was also fractionated by similar chromatographic procedures to furnish 1 (13 mg) and 36 (323 mg). Additional samples of 1 and 36 were obtained by repetition of these fermentation/isolation procedures.

(+)-(4S)-Oxazolidinone (+)-9. A flame-dried 100 mL two-necked flask was charged with (4S)-(-)-4-isopropyl-2-oxazolidinone (1.28 g, 9.90 mmol) and dry THF (15 mL) under Ar. To the solution was added n-BuLi (7.22 mL, 1.51 M in n-hexane, 10.9 mmol) dropwise at -78 °C. After stirring for 10 min at -78 °C, n-propionyl chloride (1.04 mL, 11.9 mmol) was added, and the reaction mixture was warmed to room temperature. After stirring for 2.5 h at room temperature, aqueous NH₄Cl was added. The resulting mixture was extracted with CH₂Cl₂ eight times. The combined organic extracts were washed with brine, dried over MgSO₄, filtered, and concentrated under reduced pressure to give a crude material, which was purified by flash column chromatography (hexanes-EtOAc = 3:1) to afford (+)-9 (1.75 g, 9.43 mmol, 95%) as a pale yellow oil. $R_f = 0.38$ (hexanes-EtOAc = 5:1); $[\alpha]_D^{30}$ +90 (c 1.42, CHCl₃); IR (neat, cm⁻¹): 2966,

1785, 1700, 1389, 1375, 1248, 1205, 1073, 1026, 758, 700; 1 H NMR (400 MHz, CDCl₃): δ 4.43 (ddd, J = 8.4, 4.0, 3.6 Hz, 1H), 4.27 (dd, J = 9.2, 8.4 Hz, 1H), 4.21 (dd, J = 9.2, 3.6 Hz, 1H), 3.04–2.84 (m, 2H), 2.44–2.30 (m, 1H), 1.17 (t, J = 7.2 Hz, 1H), 0.92 (d, J = 6.8 Hz, 3H), 0.88 (d, J = 6.8 Hz, 3H); 13 C NMR (100 MHz, CDCl₃): δ 174.0, 154.1, 63.3, 58.3, 29.0, 28.3, 17.9, 14.6, 8.4; HRMS (ESI) m/z: calcd for C₉H₁₅NO₃Na 208.0944 [M + Na⁺], found 208.0937.

(-)-(4R)-Oxazolidinone (-)-9. A flame-dried 50 mL two-necked flask was charged with (4R)-(-)-4-isopropyl-2-oxazolidinone (1.29 g, 10.0 mmol) and dry THF (15 mL) under Ar. To the solution was added n-BuLi (6.67 mL, 1.65 M in n-hexane, 11.0 mmol) at −78 °C. After stirring for 10 min at -78 °C, propionyl chloride (1.05 mL, 12.0 mmol) was added, and the reaction mixture was warmed to room temperature. After stirring for 3 h at room temperature, aqueous NH₄Cl was added to the reaction mixture. The resulting mixture was extracted with CH2Cl2 eight times. The combined organic extracts were dried over MgSO₄, filtered, and concentrated under reduced pressure to give a crude material, which was purified by flash column chromatography (hexanes-EtOAc = 3:1) to afford oxazolidinone (-)-9 (1.77 g, 9.53 mmol, 95%) as a pale yellow oil. $R_f = 0.80$ (hexanes-EtOAc = 1:1); $[\alpha]_D^{24}$ -81.6 (c 1.75, CHCl₃); IR (neat, cm⁻¹): 2966, 1784, 1703, 1389, 1375, 1248, 1208, 1072, 1026, 773, 700; ¹H NMR (400 MHz, CDCl₃): δ 4.47–4.40 (m, 1H), 4.27 (dd, J = 8.8, 8.4 Hz, 1H), 4.21 (dd, J = 8.8, 3.6 Hz, 1H), 3.04–2.84 (m, 2H), 2.45–2.33 (m, 1H), 1.17 (dd, I = 7.2, 5.6 Hz, 3H), 0.92 (d, I = 6.8 Hz, 3H), 0.88 (d, J = 6.8 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃): δ 173.9, 154.0, 63.3, 58.3, 29.0, 28.3, 17.8, 14.5, 8.3; LRMS (EI) m/z: 185 [M⁺]; HRMS (ESI) m/z: calcd for $C_9H_{15}NO_3Na$ 208.0944 [M + Na⁺], found 208.0938.

(-)-(2R,3S,4S)-Aldol Product 11a.7 A flame-dried 50 mL twonecked flask was charged with oxazolidinone (-)-9 (300 mg, 1.62 mmol) and dry CH₂Cl₂ (4.4 mL) under Ar. To the solution was added n-Bu₂BOTf (4.0 mL, 0.75 M in CH₂Cl₂, 3.0 mmol) at −78 °C. After stirring for 10 min, Et₃N (553 µL, 3.97 mmol) was added dropwise. The solution was stirred for 1 h at -78 °C and for 100 min at 0 °C, then recooled to -78 °C. To the resulting mixture was slowly added a solution of (S)-2-methylbutanal ((+)-10) (1.17 mL, 16.2 mmol) in dry CH₂Cl₂ (1.2 mL). After stirring for 2 h at -78 °C and for 30 min at 0 °C, the reaction was quenched with pH 6.86 phosphate buffer (5.0 mL) and MeOH (5.0 mL), followed by 34% H₂O₂ (2.4 mL). The mixture was vigorously stirred at 0 °C for 30 min. The aqueous layer was extracted with CH2Cl2 three times. The combined organic extracts were washed with NaHCO3, dried over MgSO4, filtered, and concentrated under reduced pressure to give a crude material, which was purified by flash column chromatography (hexanes-EtOAc = 7:3) to afford aldol product 11a (275 mg, 1.01 mmol, 63%) as a white solid. $R_f = 0.31$ (hexanes-EtOAc = 3:1); m.p.: 98–101 °C (hexanes-CHCl₃, white needle); $[\alpha]_{D}^{24}$ -70 (c 0.80, CH₂Cl₂); IR (ATR, cm⁻¹): 3463, 2963, 1764, 1684, 1387, 1372, 1238, 1206, 777, 705, 640; ¹H NMR (400 MHz, CDCl₃): δ 4.47 (dt, J = 8.4, 3.2 Hz, 1H), 4.29 (dd, J = 9.6, 8.4 Hz, 1H), 4.22 (dd, J = 9.6, 3.2 Hz, 1H), 4.01–3.93 (m, 1H), 3.63– 3.58 (m, 1H), 3.11-3.07 (m, 1H), 2.42-2.28 (m, 1H), 1.87-1.73 (m, 1H), 1.55-1.50 (m, 1H), 1.24-1.12 (m, 1H), 1.23 (d, J = 7.2 Hz, 3H), 0.93 (d, J = 7.6 Hz, 3H), 0.91 (t, J = 8.0 Hz, 3H), 0.89 (d, J = 6.8Hz, 3H), 0.85 (d, J = 6.8 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃): δ 178.4, 153.4, 74.5, 63.3, 58.2, 39.3, 36.7, 28.3, 25.1, 17.9, 14.70, 14.67, 10.8, 10.0; LRMS (EI) m/z: 272 [M + H⁺]; HRMS (ESI) m/z: calcd for $C_{14}H_{25}NO_4Na$ 294.1676 [M + Na⁺], found 294.1679.

The Structure of 11a Was Confirmed by the Following Procedure. A screw cap test tube was charged with 11a (60 mg, 0.22 mmol) and dry Et₂O (0.8 mL). To the solution was added LiBH₄ (10 mg, 0.442 mmol) at -42 °C. After stirring for 14 h at room temperature, aqueous NH₄Cl was added at 0 °C. The resulting mixture was extracted with CH₂Cl₂ three times. The combined organic extracts were washed with brine, dried over NaSO₄, filtered, and concentrated under reduced pressure to give a crude material, which was purified by silica gel column chromatography (hexanes-EtOAc = 3:1) to afford S1 (20.1 mg, 0.137 mmol, 62%) as a white solid. $R_f = 0.36$ (hexanes-EtOAc = 1:1); m.p.: 80–81 °C (hexanes-EtOAc, white needle); $[\alpha]_{D}^{26}$ -6.1 (c 0.14, CHCl₃); IR (neat, cm⁻¹): 3353, 2964, 2933, 2877, 1748,

1463, 1384, 1144, 1074, 1032, 973; $^{1}\mathrm{H}$ NMR (400 MHz, CDCl_3): δ 3.82–3.68 (m, 2H), 3.56–3.50 (m, 1H), 2.00 (d, J = 4.4 Hz, 2H), 1.90–1.80 (m, 1H), 1.80–1.68 (m, 1H), 1.28–1.10 (m, 1H), 0.98 (d, J = 8.8 Hz, 3H), 0.93 (t, J = 7.6 Hz, 3H), 0.83 (d, J = 6.8 Hz, 2H); $^{13}\mathrm{C}$ NMR (150 MHz, CDCl_3): δ 77.4, 67.9, 37.7, 36.0, 25.2, 14.9, 10.8, 8.7. Spectral data matched the reported data of \$1.28

(+)-(2S,3R,4S)-Aldol Product 11b.7 A flame-dried 50 mL twonecked flask was charged with oxazolidinone (+)-9 (300 mg, 1.62 mmol) and dry CH₂Cl₂ (4.4 mL) under Ar. To the solution was added n-Bu₂BOTf (4.0 mL, 0.75 M in CH₂Cl₂, 3.0 mmol) at −78 °C. After stirring for 10 min, Et₃N (553 µL, 3.97 mmol) was added dropwise. The solution was stirred for 1 h at -78 °C and for 100 min at 0 °C, then recooled to -78 °C. To the resulting mixture was slowly added a solution of (S)-2-methylbutanal ((+)-10) (1.17 mL, 16.2 mmol) in dry CH₂Cl₂ (1.2 mL). After stirring for 2 h at -78 °C and for 1 h at 0 °C, the reaction was quenched with pH 6.86 phosphate buffer (5.0 mL) and MeOH (5.0 mL), followed by 34% H₂O₂ (2.4 mL). The mixture was vigorously stirred at 0 $^{\circ}\text{C}$ for 30 min. The aqueous layer was extracted with CH₂Cl₂ three times. The combined organic extracts were washed with NaHCO3, dried over MgSO4, filtered, and concentrated under reduced pressure to give a crude material, which was purified by flash column chromatography (hexanes-EtOAc = 7:3) to afford aldol product 11b (364 mg, 1.34 mmol, 83%) as a white solid. $R_f = 0.23$ (hexanes-EtOAc = 3:1); m.p.: 80-81 °C (hexanes-CHCl₃, white needle); $[\alpha]_{\rm D}^{23}$ +74 (*c* 0.80, CH₂Cl₂); IR (ATR, cm⁻¹): 3531, 2963, 1766, 1681, 1381, 1209, 988, 957, 780, 706; ¹H NMR (400 MHz, CDCl₃): δ 4.47 (ddd, J = 8.0, 3.6, 3.6 Hz, 1H), 4.29 (dd, J= 9.6, 8.0 Hz, 1H), 4.22 (dd, J = 9.6, 3.6 Hz, 1H), 4.00 (qd, J = 7.2, 3.2)Hz, 1H), 3.67 (ddd, J = 7.2, 4.0, 3.2 Hz, 1H), 2.75 (d, J = 4.0 Hz, 1H), 2.35 (m, 1H₁), 1.55–1.40 (m, 2H), 1.26 (d, J = 7.2 Hz, 3H), 1.20– 1.05 (m, 1H), 0.98 (d, J = 6.8 Hz, 3H), 0.92 (d, J = 6.8 Hz, 3H), 0.90 (t, J = 7.2 Hz, 3H), 0.89 (d, J = 6.8 H, 3H); 13 C NMR (100 MHz, CDCl₂): δ 177.9, 153.4, 74.7, 63.3, 58.2, 39.7, 37.0, 28.3, 25.6, 17.9, 14.7, 14.6, 11.5, 11.2; LRMS (EI) m/z: 272 [M + H⁺]; HRMS (ESI) m/z: calcd for C₁₄H₂₅NO₄Na 294.1676 [M + Na⁺], found 294.1674. Spectral data matched the reported data.

(+)-(2R,3R,4S)-Aldol Product 11c. 12 A flame-dried 30 mL twonecked flask was charged with oxazolidinone (+)-9 (185 mg, 1.00 mmol) and dry CH₂Cl₂ (2.0 mL) under Ar. To the solution were added DIPEA (200 μ L, 1.15 mmol) and n-Bu₂BOTf (330 mg, 1.20 mmol) at 0 °C. After stirring for 75 min, the solution was cooled to -78 °C. In a separate flame-dried 30 mL two-necked flask, (S)-2methylbutanal ((+)-10) (108 μ L, 1.50 mmol) was placed, to which was added a solution of SnCl₄ (88 µL, 0.75 mmol) in 2.0 mL of dry CH₂Cl₂. The resulting mixture was stirred for 5 min and transferred to the enol borate solution via cannula with aid of additional CH₂Cl₂ (1.0 mL). After stirring for 3 h, the reaction was quenched with MeOH-34% H₂O₂ aq. (5:1, 6.0 mL). Stirring was continued at -78 °C for another 10 min, after which time the solution was warmed to 0 $^{\circ}\text{C}$ and stirred for 30 min. To the solution was added water, and the aqueous layer of the resulting mixture was extracted twice with ether. The combined organic extracts were washed with saturated NaHCO3 and brine, dried over MgSO₄, filtered, and concentrated under reduced pressure to give a crude material, which was purified by flash column chromatography (hexanes-EtOAc = 3:1) to afford aldol product 11c (127 mg, 0.468 mmol, 47%) as a white solid. $R_f = 0.20$ (hexanes-EtOAc = 3:1); $[\alpha]_D^{28}$ +64 (c 0.82, CH₂Cl₂); IR (ATR, cm⁻¹): 3463, 2961, 1757, 1702, 1371, 1221, 1208, 775, 712; ¹H NMR (400 MHz, CDCl₃): δ 4.44 (ddd, J = 8.4, 3.6, 3.6 Hz, 1H), 4.28 (dd, J = 9.2, 8.4 Hz, 1H), 4.23 (dd, J = 9.2, 3.6 Hz, 1H), 4.06 (dq, J = 8.4, 6.8 Hz, 1H), 3.62 (ddd, J = 10.0, 8.4, 3.6 Hz, 1H), 2.51 (d, J = 10.0 Hz, 1H), 2.48-2.37 (m, 1H), 1.60-1.41 (m, 2H), 1.37-1.24 (m, 1H), 1.13 (d, J = 7.6Hz, 3H), 0.98–0.88 (m, 12H); 13 C NMR (100 MHz, CDCl₃): δ 177.2, 154.7, 77.6, 63.4, 59.0, 40.4, 37.0, 28.5, 26.7, 18.0, 14.7, 14.6, 12.2, 11.8; LRMS (EI) m/z: 271 [M⁺]; HRMS (ESI) m/z: calcd for $C_{14}H_{25}NO_4Na$ 294.1676 [M + Na⁺], found 294.1681.

(–)-(2R,3S,4S)-Oxazolidinone 11d. A flame-dried 30 mL two-necked flask was charged with oxazolidinone (–)-9 (370 mg, 2.00 mmol) and dry CH₂Cl₂ (4.0 mL) under Ar. To the colorless solution were added DIPEA (400 μ L, 2.30 mmol) and n-Bu₂BOTf (660 mg,

2.40 mmol) at 0 °C. After stirring for 50 min, the solution was cooled to -78 °C. In a separate flame-dried 30 mL two-necked flask, (S)-2methylbutanal ((+)-10) (108 μ L, 1.50 mmol) was placed, to which was added a solution of SnCl₄ (176 µL, 1.50 mmol) in 4.0 mL of dry CH₂Cl₂. After stirring for 5 min, the solution was transferred to the enol borate solution via cannula with aid of additional CH₂Cl₂ (2.0 mL). After stirring for 6 h, the reaction was quenched with MeOH-34% H₂O₂ aq. (5:1, 12 mL). Stirring was continued at -78 °C for another 10 min, after which time the solution was warmed to 0 °C and stirred for 30 min. To the solution was added water, and the aqueous layer was extracted twice with ether. The combined organic extracts were washed with saturated NaHCO3 and brine, dried over MgSO4, filtered, and concentrated under reduced pressure to give a crude material, which was purified by flash column chromatography (hexanes-EtOAc = 3:1) to afford aldol product 11d (193 mg, 0.713 mmol, 36%) as a white solid. $R_f = 0.21$ (hexanes-EtOAc = 3:1); $[\alpha]_D^{28}$ -60 (c 0.76, CH₂Cl₂); IR (ATR, cm⁻¹): 3506, 2963, 1769, 1692, 1384, 1202, 710, 629; ¹H NMR (400 MHz, CDCl₃): δ 4.45 (ddd, J = 7.2, 3.6, 3.6 Hz, 1H), 4.28 (dd, J = 9.2, 7.2 Hz, 1H), 4.22 (dd, J = 9.2, 3.6 Hz, 1H), 4.13 (qd, J = 7.2, 6.4 Hz, 1H), 3.44 (ddd, J = 10.0, 6.4, 6.4 Hz, 1H), 2.76 (d, J = 10.0 H, 1H), 2.46-2.34 (m, 1H), 1.76-1.62 (m, 1H), 1.56-1.46 (m, 1H), 1.21 (d, I = 7.2 Hz, 3H), 1.28-1.12 (m, 1H), 0.97–0.88 (m, 12H); 13 C NMR (100 MHz, CDCl₃): δ 177.6, 154.2, 79.5, 63.2, 58.8, 39.6, 38.0, 28.5, 23.1, 17.9, 16.0, 15.2, 14.6, 11.5; LRMS (EI) m/z: 272 [M + H⁺]; HRMS (ESI) m/z: calcd for C₁₄H₂₅NO₄Na 294.1676 [M + Na⁺], found 294.1678.

The Structure of 11d Was Confirmed by the Following Procedure. A screw cap test tube was charged with 11d (4.5 mg, 17 μ mol), dry CH₂Cl₂ (55 μ L) and 2,6-lutidine (5.5 μ L, 47.6 μ mol). To the solution was added TBSOTf (5.8 μ L, 0.442 mmol) at -42 °C. After stirring for 14 h at room temperature, the reaction mixture was quenched with H2O. The resulting mixture was extracted with CH2Cl2 three times. The combined organic extracts were washed with saturated aqueous NaHCO3, brine, dried over Na2SO4, filtered, and concentrated under reduced. The residue was purified by preparative TLC (hexanes-EtOAc = 5:1) to afford silyl ether S2 as a pale yellow oil (3.1 mg, 11 μ mol, 65%). $R_f = 0.53$ (hexanes-EtOAc = 3:1). $[\alpha]_D^{26}$ -12.9 (c 1.34, CHCl₃); IR (neat, cm⁻¹): 2960, 2928, 2880, 2855, 1782, 1699, 1387, 1254, 1202, 1057, 837; ¹H NMR (400 MHz, CDCl₃): δ 4.46–4.38 (m, 1H), 4.30–4.15 (m, 2H), 4.10–4.00 (m, 1H), 2.44-2.30 (m, 1H), 1.63-1.44 (m, 3H), 1.22-1.10 (m, 1H), 1.12 (d, J = 6.4 Hz, 3H), 0.94-0.89 (m, 12H), 0.89-0.84 (m, 9H),0.09 (s, 3H), 0.04 (s, 3H); 13 C NMR (100 MHz, CDCl₃): δ 175.3, 153.7, 62.6, 58.8, 43.3, 38.9, 28.3, 28.2, 26.1, 24.4, 18.3, 18.2, 16.3, 14.2, 13.7, 12.4, -4.2, -4.7 (two signals are missing due to overlap); HRMS (ESI⁺) m/z: calcd for C₂₀H₃₉NO₄SiNa 408.2541 [M + Na⁺], found 408.2534. A screw cap test tube was charged with silyl ether S2 (3.1 mg, 11 μ mol), dry MeOH (2.2 μ L, 55 μ mol), and dry THF (55 μ L). To the solution was added LiBH₄ (1.2 mg, 55 μ mol) at 0 °C. After stirring for 10 h at room temperature, 1 M aqueous NaOH was added at 0 °C. then warmed to room temperature. The resulting mixture was extracted with CH2Cl2 three times. The combined organic extracts were washed with brine, dried over NaSO4, filtered, and concentrated under reduced pressure to give a crude material, which was purified by silica gel column chromatography (hexanes-EtOAc = 5:1) to afford alcohol S3 (0.8 mg, 11 μ mol, 28%) as a pale yellow oil. $R_f = 0.63$ (hexanes-ethyl acetate = 3:1); $[\alpha]_D^{26}$ +6.3 (c 0.52, CHCl₃); IR (neat, cm⁻¹): 3483, 2959, 2930, 2857, 1787, 1706, 1464, 1387, 1254, 1053, 1026, 836; ¹H NMR (400 MHz, CDCl₃): δ 3.70–3.62 (m, 1H), 3.62-3.50 (m, 2H), 2.74 (br s, 1H), 1.92-1.80 (m, 1H), 1.62-1.44 (m, 2H), 1.22-1.04 (m, 1H), 0.98 (d, J = 7.2 Hz, 3H), 0.95-0.85 (m, 2H)15H), 0.11 (s, 3H), 0.09 (s, 3H); 13 C NMR (150 MHz, CDCl₃): δ 81.7, 66.4, 41.0, 36.4, 26.0, 25.3, 18.2, 16.7, 14.8, 12.3, -4.23 (three signals are missing due to overlap). Spectral data matched the reported data of S3.29

(–)-(2R,3S,4S)-β-Hydroxy Ester **8a**. ¹⁰ A screw cap test tube was charged with aldol product **11a** (15 mg, 55 μ mol) and anhydrous MeOH (120 μ L) at 0 °C. To the solution was added sodium methoxide (3.5 mg, 64 μ mol). After stirring for 90 min at 0 °C, the reaction was quenched with aqueous NH₄Cl. The mixture was

extracted with EtOAc three times. The combined organic extracts were dried over MgSO₄, filtered, and concentrated under reduced pressure to give a crude material, which was purified by flash column chromatography (hexanes-EtOAc = 5:1) to afford β-hydroxy ester 8a (8.8 mg, 51 μmol, 91%) as a colorless oil. R_f = 0.38 (hexanes-EtOAc = 3:1); $[\alpha]_D^{24}$ –4.1 (c 0.60, CH₂Cl₂); IR (neat, cm⁻¹): 3507, 2964, 1735, 1720, 1458, 1202, 1056, 996, 968; ¹H NMR (400 MHz, CDCl₃): δ 3.71 (s, 3H), 3.67 (ddd, J = 8.0, 4.0, 3.2 Hz, 1H), 2.68 (qd, J = 7.2, 3.2 Hz, 1H), 2.48 (d, J = 4.0 Hz, 1H), 1.77 (dqd, J = 15.2, 7.6, 2.8 Hz, 1H), 1.51–1.41 (m, 1H), 1.17 (d, J = 7.2 Hz, 3H), 1.24–1.12 (m, 1H), 0.91 (t, J = 7.6 Hz, 3H), 0.83 (d, J = 7.2 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃): δ 177.1, 75.1, 51.7, 41.4, 36.9, 24.8, 14.9, 10.9, 9.6; LRMS (EI) m/z: 175 [M + H⁺]; HRMS (ESI) m/z: calcd for $C_9H_{18}O_3Na$ 197.1148 [M + Na⁺], found 197.1143.

C₉H₁₈O₃Na 197.1148 [M + Na⁺], found 197.1143. (–)-(2S,3R,4S)- β -Hydroxy Ester **8b**.¹⁰ A screw cap test tube was charged with aldol product 11b (10 mg, 37 µmol) and anhydrous MeOH (80 μ L) at 0 $^{\circ}$ C. To the solution was added sodium methoxide (2.0 mg, 43 μ mol). After stirring for 20 min at 0 °C, the reaction was quenched with aqueous NH₄Cl. The mixture was extracted with EtOAc three times. The combined organic extracts were dried over MgSO₄, filtered, and concentrated under reduced pressure to give a crude material, which was purified by flash column chromatography (hexanes-EtOAc = 5:1) to afford β -hydroxy ester **8b** (5.2 mg, 30 μ mol, 81%) as a colorless oil. $R_f = 0.36$ (hexanes-EtOAc = 3:1); $[\alpha]_D^{24} - 1.2$ (c 0.53. CH₂Cl₂); IR (neat, cm⁻¹): 3504, 2964, 1737, 1720, 1459, 1261, 1200, 1169; ¹H NMR (400 MHz, CDCl₃): δ 3.72–3.65 (m, 1H), 3.70 (s, 3H), 2.68 (qd, *J* = 7.2, 5.2 Hz, 1H), 2.15 (d, *J* = 4.8 Hz, 1H), 1.52– 1.34 (m, 2H), 1.22–1.10 (m, 1H), 1.21 (d, J = 7.2 Hz, 3H), 0.96 (d, J = 6.8 Hz, 3H), 0.90 (t, J = 7.2 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃): δ 176.6, 75.1, 51.7, 42.2, 37.1, 25.9, 13.9, 11.5, 11.1; LRMS (EI) m/z: 174 [M⁺]; HRMS (ESI) m/z: calcd for C₉H₁₈O₃Na 197.1148 [M + Na⁺], found 197.1149. Spectral data matched the reported data.7

(–)-(2R,3R,4S)-β-Hydroxy Ester **8c**. ¹⁰ A screw cap test tube was charged with aldol product 11c (10 mg, 37 μ mol) and anhydrous MeOH (80 μ L) at 0 °C. To the solution was added sodium methoxide (2.0 mg, 43 μ mol). After stirring for 20 min at 0 °C, the reaction was quenched with aqueous NH₄Cl, and the mixture was extracted with EtOAc three times. The combined organic extracts were dried over MgSO₄, filtered, and concentrated under reduced pressure to give a crude material, which was purified by flash column chromatography (hexanes-EtOAc = 3:1) to afford β -hydroxy ester 8c (5.3 mg, 30 μ mol, 82%) as a colorless oil. $R_f = 0.33$ (hexanes-EtOAc = 3:1); $[\alpha]_D^{26} - 9.3$ (c 0.45, CH₂Cl₂); IR (neat, cm⁻¹): 3523, 2963, 1738, 1721, 1458, 1260, 1198, 1172, 992, 774; ¹H NMR (400 MHz, CDCl₃): δ 3.72 (s, 3H), 3.62 (ddd, J = 8.0, 6.4, 3.6 Hz, 1H), 2.65 (dq, J = 8.0, 7.2 Hz, 1H), 2.37(d, J = 6.4 Hz, 1H), 1.54 - 1.40 (m, 2H), 1.36 - 1.23 (m, 1H), 1.16 (d, J= 7.2 Hz, 3H), 0.92 (t, J = 7.6 Hz, 3H), 0.88 (d, J = 6.8 Hz, 3H); 13 C NMR (100 MHz, CDCl₃): δ 177.0, 75.7, 51.8, 43.0, 36.8, 26.6, 14.4, 12.4, 11.7; LRMS (EI) m/z: 175 [M + H⁺]; HRMS (ESI) m/z: calcd for $C_9H_{18}O_3Na$ 197.1148 [M + Na⁺], found 197.1139. Spectral data matched the reported data.

(+)-(2R,3S,4S)-β-Hydroxy Ester 8d. 10 A screw cap test tube was charged with aldol product 11d (10 mg, 37 μ mol) and anhydrous MeOH (80 μ L) at 0 °C. To the solution was added sodium methoxide (2.0 mg, 43 μ mol). After stirring for 20 min at 0 °C, the reaction was quenched with aqueous NH₄Cl, and the mixture was extracted with EtOAc three times. The combined organic extracts were dried over MgSO₄, filtered, and concentrated under reduced pressure to give a crude material, which was purified by flash column chromatography (hexanes-EtOAc = 3:1) to afford β-hydroxy ester 8d (4.1 mg, 24 μ mol, 64%) as a colorless oil. $R_f = 0.33$ (hexanes-EtOAc = 3:1); $[\alpha]_D^{25} + 4.8$ (c 0.51, CH₂Cl₂); IR (neat, cm⁻¹): 3522, 2963, 1734, 1719, 1458, 1260, 1199, 1172, 1044, 997; ¹H NMR (400 MHz, CDCl₃): δ 3.71 (s, 3H), 3.38 (ddd, J = 8.4, 6.4, 5.6 Hz, 1H), 2.73 (qd, J = 7.2, 5.6 Hz, 1H), 2.56(d, J = 8.4 Hz, 1H), 1.69-1.58 (m, 1H), 1.53-1.41 (m, 1H), 1.24 (d, J= 7.2 Hz, 3H), 1.22-1.12 (m, 1H), 0.92 (d, J = 6.8 Hz, 3H), 0.90 (t, J= 7.6 Hz, 3H); 13 C NMR (100 MHz, CDCl₃): δ 176.9, 77.9, 51.7, 41.9, 38.2, 23.6, 15.8, 15.1, 11.4; LRMS (EI) m/z: 175 [M + H⁺]; HRMS (ESI) m/z: calcd for C₉H₁₈O₃Na 197.1148 [M + Na⁺], found 197.1147.

(-)-(2R,3S,4S)-THP-Protected Aldol Product (-)-12. A screw cap test tube was charged with aldol product (-)-11a (250 mg, 0.921 mmol) and dry CH₂Cl₂ (3.0 mL). To the solution were added DHP (168 μ L, 1.84 mmol) and TsOH·H₂O (1.8 mg, 9.2 μ mol). After stirring for 3 h at room temperature, the reaction was quenched with H₂O. The aqueous layer was extracted with CH₂Cl₂ two times. The combined organic extracts were dried over MgSO₄, filtered, and concentrated under reduced pressure to give a crude material, which was purified by flash column chromatography (hexanes-EtOAc = 3:1) to afford THP-protected β -hydroxy ester (-)-12 (295 mg, 0.830 mmol, 90%) as a pale yellow oil. $R_f = 0.26$ (hexanes-EtOAc = 3:1). The physical data of diastereomeric mixture (-)-12: $[\alpha]_D^{26}$ -70.7 (c 1.04, CHCl₃); IR (neat, cm⁻¹): 2962, 1780, 1700, 1384, 1227, 1202, 903, 753; ¹H NMR (400 MHz, CDCl₃): δ 4.57–4.52 (m, 0.5H), 4.45-4.40 (m, 0.5H), 4.40-4.30 (m, 1H), 4.28-4.20 (m, 1H), 4.22-4.18 (m, 1H), 4.10-4.00 (m, 0.5H), 4.01-3.89 (m, 0.5H), 3.88-3.81 (m, 1H), 3.81–3.76 (m, 1H), 3.48–3.34 (m, 1H), 2.52–2.34 (m, 1H), 1.85-1.66 (m, 2H), 1.63-1.40 (m, 6H), 1.21 (dd, J = 6.8, 3.2 Hz, 3H), 1.13-1.01 (m, 1H) 0.97 (d, J = 7.2 Hz, 3H), 0.95-0.85 (m, 9H); 13 C NMR (100 MHz, CDCl₃): δ 176.4, 175.0, 154.1, 153.5, 101.3, 100.3, 82.0, 81.9, 64.3, 63.34, 63.31, 63.2, 59.3, 58.9, 40.7, 40.0, 39.2, 38.3, 30.90, 30.85, 28.4, 25.4, 25.3, 25.0, 24.8, 20.8, 20.2, 18.1, 18.0, 15.2, 14.8, 14.6, 13.1, 12.0, 11.7, 11.1 (two signals are missing due to overlap); HRMS (ESI) m/z: calcd for $C_{19}H_{33}NO_5Na$ 378.2251 [M + Na⁺], found 378.2253.

(-)-(2R,3S,4S)-THP-Protected β -Hydroxy Carboxylic Acid (-)-13. A screw cap test tube was charged with (-)-12 (9.1 mg, 26 μ mol), dry THF (75 μ L), and H₂O (22 μ L). To the solution were added H₂O₂ (34% w/w solution in H₂O, 20 µL, 0.18 mmol) and LiOH·H₂O (1.6 mg, 38 μ mol) at 0 °C. The reaction mixture was warmed to at room temperature and stirred for 24 h. The reaction was quenched with saturated aqueous Na₂S₂O₃ and 0.1 M HCl aq. The aqueous layer was extracted with EtOAc 10 times. The combined organic extracts were dried over Na2SO4, filtered, and concentrated under reduced pressure to give a crude material, which was purified by flash column chromatography (hexanes-EtOAc = 5:1) to afford THP-protected β hydroxy carboxylic acid (-)-13 (5.1 mg, 21 μ mol, 82%) as a pale yellow oil. $R_f = 0.13$ (hexanes-EtOAc = 3:1). The physical data of diastereomeric mixture (–)-13: $[\alpha]_{\rm D}^{28}$ –19 (c 0.80, CHCl₃); IR (neat, cm⁻¹): 2942, 1708, 1457, 1131, 1077, 1034, 813, 669; ¹H NMR (400 MHz, CDCl₃): δ 4.62–4.58 (m, 0.5H), 4.56–4.51 (m, 0.5H), 3.97– 3.84 (m, 1.5H), 3.78 (dd, J = 6.0, 4.4 Hz, 0.5H), 3.52 - 3.41 (m, 1H),2.86-2.77 (m, 0.5H), 2.76-2.68 (m, 0.5H), 1.85-1.70 (m, 2H), 1.70-1.45 (m, 6H), 1.21 (d, J = 7.2 Hz, 1.5H), 1.16 (d, J = 6.8 Hz, 1.5H), 1.20–1.15 (m, 1H), 0.95 (d, J = 7.2 Hz, 1.5H), 0.91 (d, J = 6.8 Hz, 1.5H), 0.89 (t, J = 7.2 Hz, 3H); 13 C NMR (100 MHz, CDCl₃): δ 181.5, 179.8, 101.7, 100.0, 84.0, 82.1, 63.9, 63.4, 41.8, 41.3, 38.5, 37.3, 31.03, 31.00, 25.4, 25.1, 25.0, 24.5, 20.7, 20.2, 15.6, 15.4, 11.7, 11.5, 11.2 (one signal is missing due to overlap); HRMS (ESI) m/z: calcd for $C_{13}H_{24}O_4Na$ 267.1567 [M + Na⁺], found 267.1560. (+)-(25,3R,4R)-Aldol Product 11e.⁷ A flame-dried 20 mL two-

necked flask was charged with oxazolidinone (+)-9 (200 mg, 1.08 mmol) and dry CH₂Cl₂ (1.9 mL) under Ar. To the solution was added n-Bu₂BOTf (1.2 mL, 1.00 M in CH₂Cl₂, 1.19 mmol) at −78 °C. After stirring for 10 min, Et₃N (210 μ L, 1.51 mmol) was added dropwise. The solution was stirred for 1 h at −78 °C and for 1 h at 0 °C, then recooled to -78 °C. To the resulting mixture was slowly added a solution of (R)-2-methylbutanal $((-)-10)^9$ (110 mg, 1.28 mmol) in dry CH_2Cl_2 (868 μ L). After stirring for 30 min at -78 °C and for 2 h at 0 °C, the reaction was quenched with pH 6.86 phosphate buffer (868 μ L) and MeOH (868 μ L), followed by 34% H₂O₂ (34% w/w solution in H_2O , 520 μL). The mixture was vigorously stirred at 0 °C for 1.5 h. The aqueous layer was extracted with CH2Cl2 three times. The combined organic extracts were washed with brine and dried over MgSO₄, filtered, and concentrated under reduced pressure to give a crude material, which was purified by flash column chromatography (hexanes-EtOAc = 3:1) to afford aldol product 11e (154 mg, 0.567 mmol, 53%) as a white solid. $R_f = 0.19$ (hexanes-EtOAc = 3:1); $[\alpha]_D^{26}$

+61 (c 0.80, CH₂Cl₂); IR (ATR, cm⁻¹): 3464, 2963, 1765, 1685, 1387, 1372, 1238, 1205, 704, 640; ¹H NMR (400 MHz, CDCl₃): δ 4.47 (ddd, J = 9.2, 8.4, 3.2 Hz, 1H), 4.29 (dd, J = 9.2, 8.4 Hz, 1H), 4.22 (dd, J = 9.2, 3.2 Hz, 1H), 3.97 (qd, J = 7.2, 2.4 Hz, 1H), 3.61 (ddd, J = 9.2, 8.4 Hz, 1H), 3.10 (d, J = 2.8 Hz, 1H), 2.40–2.30 (m, 1H), 1.86–1.74 (m, 1H), 1.55–1.40 (m, 1H), 1.23 (d, J = 7.2 Hz, 3H), 1.27–1.24 (m, 1H), 0.93 (d, J = 7.6 Hz, 3H), 0.91 (t, J = 7.2 Hz, 3H), 0.89 (d, J = 6.8 Hz, 3H), 0.85 (d, J = 6.8 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃): δ 178.4, 153.4, 74.5, 63.3, 58.2, 39.3, 36.7, 28.3, 25.2, 17.9, 14.70, 14.67, 10.8, 10.0; HRMS (ESI) m/z: calcd for C₁₄H₂₅NO₄Na 294.1676 [M + Na⁺], found 294.1688. Spectral data matched the reported data. ¹³

(+)-(2S,3R,4R)-THP-Protected Aldol Product (+)-12. A screw cap test tube was charged with aldol product 11e (30 mg, 0.11 mmol) and dry CH₂Cl₂ (349 μ L). To the solution were added DHP (20 μ L, 0.22 mmol) and TsOH·H₂O (0.21 mg, 1.1 μ mol). After stirring for 2 h at room temperature, the reaction was quenched with H₂O. The aqueous layer was extracted with CH2Cl2 five times. The combined organic extracts were dried over Na2SO4, filtered, and concentrated under reduced pressure to give a crude material, which was purified by flash column chromatography (hexanes-EtOAc = 5:1) to afford (+)-12 (29.8 mg, 83.8 μ mol, 76%) as a pale yellow oil. $R_f = 0.35$ (hexanes-EtOAc = 3:1). The physical data of diastereomeric mixture (+)-12 is following: $[\alpha]_D^{28}$ +68 (c 0.52, CHCl₃); IR (neat, cm⁻¹): 2962, 1780, 1700, 1385, 1228, 1202, 1130, 1032; 1 H NMR (400 MHz, CDCl₃): δ 4.57-4.52 (m, 0.5H), 4.45-4.40 (m, 0.5H), 4.40-4.30 (m, 1H), 4.28-4.20 (m, 1H), 4.22-4.18 (m, 1H), 4.10-4.00 (m, 0.5H), 4.00-3.88 (m, 0.5H), 3.86-3.76 (m, 2H), 3.48-3.32 (m, 1H), 2.52-2.32 (m, 1H), 1.86-1.76 (m, 2H), 1.65-1.38 (m, 6H), 1.21 (dd, <math>J = 6.8,3.6 Hz, 3H), 1.13-1.00 (m, 1H), 0.98-0.95 (m, 3H), 0.94-0.83 (m, 9H); 13 C NMR (100 MHz, CDCl₃): δ 176.5, 175.0, 154.1, 153.6, 101.3, 100.3, 82.1, 81.9, 64.3, 63.35, 63.31, 63.2, 59.3, 58.9, 40.7, 40.0, 39.3, 38.3, 30.91, 30.87, 28.4, 25.4, 25.3, 25.0, 24.8, 20.8, 20.2, 18.1, 18.0, 15.2, 14.8, 14.6, 13.2, 12.0, 11.7, 11.1 (two signals are missing due to overlap); HRMS (ESI) m/z: calcd for C₁₉H₃₃NO₅Na 378.2251 [M + Na⁺], found 378.2253.

(+)-(2S,3R,4R)-THP-Protected β -Hydroxy Carboxylic Acid (+)-13. A screw cap test tube was charged with THP-protected aldol product (+)-12 (30 mg, 0.084 mmol), dry THF (250 μ L) and H₂O (72 μ L). To the solution were added H_2O_2 (34% w/w solution in H_2O , 60 μ L, 0.59 mmol) and LiOH·H₂O (5.3 mg, 0.13 mmol) at 0 $^{\circ}$ C. The reaction mixture was warmed to room temperature and stirred for 24 h. The reaction was quenched with saturated aqueous Na₂S₂O₃ and 0.1 M HCl aq. The aqueous layer was extracted with EtOAc six times. The combined organic extracts were dried over Na2SO4, filtered, and concentrated under reduced pressure to give a crude material, which was purified by flash column chromatography (hexanes-EtOAc = 3:1) to afford THP-protected β -hydroxy carboxylic acid (+)-13 (17.3 mg, 70.8 mmol, 84%) as a pale yellow oil. $R_f = 0.22$ (hexanes-EtOAc = 3:1). The physical data of diastereomeric mixture (+)-13 is following: $[\alpha]_{\rm D}^{28}$ +20 (c 0.59, CHCl₃); IR (neat, cm⁻¹): 2941, 1707, 1456, 1201, 1131, 1077, 1026; ¹H NMR (400 MHz, CDCl₃): δ 4.65–4.60 (m, 0.5H), 4.60-4.54 (m, 0.5H), 4.00-3.85 (m, 1.5H), 3.77-3.72 (m, 0.5H), 3.54-3.44 (m, 1H), 2.94-2.84 (m, 0.5H), 2.80-2.70 (m, 0.5H), 1.87-1.77 (m, 2H), 1.64-1.60 (m, 6H), 1.21(d, J = 7.2 Hz, 1.5H), 1.18 (d, J = 7.2 Hz, 1.5H), 1.18–1.10 (m, 1H), 0.98–0.85 (m, 6H); 13 C NMR (100 MHz, CDCl₃): δ 181.6, 180.0, 101.6, 100.0, 84.0, 82.1, 63.8, 63.4, 41.8, 41.3, 38.6, 37.4, 31.03, 31.00, 25.4, 25.1, 25.0, 24.5, 20.7, 20.2, 15.5, 15.4, 11.7, 11.5, 11.2 (one signal is missing due to overlap); HRMS (ESI) m/z: calcd for $C_{13}H_{24}O_4Na$ 267.1567 [M + Na⁺], found 267.1563.

(–)-Deacetylaranotin (5). ¹⁴ A screw cap test tube was charged with acetylaranotin (–)-1 (2.0 mg, 40 μ mol) and dry CH₂Cl₂ (100 μ L). To the solution was added 36% HCl aq. (2.5 mL) in MeOH (98 μ L). The reaction mixture was stirred at room temperature for 16.5 h. The reaction mixture was diluted with EtOAc, water, and saturated aqueous NaHCO₃. The resulting mixture was extracted with EtOAc three times. The organic layer was dried over Na₂SO₄, filtered, and concentrated under reduced pressure to give a crude material, which was purified by preparative TLC (CH₂Cl₂-acetone = 20:1) to afford

(–)-deacetylaranotin (5) (1.2 mg, 2.9 μmol, 72%) as a white solid. $R_f = 0.27$ (CH₂Cl₂-acetone = 10:1); [α]_D²⁶ –546 (ϵ 0.210, CHCl₃); IR (KBr, cm⁻¹): 3339, 1693, 1666, 1655, 1445, 1431, 1387, 1283, 1144, 1036; ¹H NMR (600 MHz, CDCl₃): δ 6.60 (ϵ , 2H), 6.27 (dd, f = 8.9, 2.4 Hz, 2H), 5.88 (ϵ , 2H), 4.81 (dd, f = 8.4, 2.4 Hz, 2H), 4.78 (d, f = 7.2 Hz, 2H), 4.62 (d, f = 7.2 Hz, 2H), 3.88 (dt, f = 16.8, 1.8 Hz, 2H), 2.92 (d, f = 16.8 Hz, 2H); ¹³C NMR (150 MHz, CDCl₃): δ 165.4, 139.0, 138.9, 109.7, 107.6, 75.4, 70.7, 68.0, 36.2; HRMS (ESI⁺) m/z: calcd for $C_{18}H_{16}N_2O_6S_2Na$ 443.0347 [M + Na⁺], found 443.0358.

(-)-Diester 14. A screw cap test tube was charged with (-)-deacetylaranotin (5) (2.30 mg, 5.47 μ mol), THP-protected β hydroxy carboxylic acid (-)-13 (5.1 mg, 20.9 μ mol), and dry CH₂Cl₂ (100 μ L). To the solution were added DMAP (2.00 mg, 16.4 μ mol) and WSCD·HCl (5.24 mg, 27.4 µmol). The reaction mixture was stirred for 5 h at room temperature. The reaction mixture was diluted with hexane and EtOAc, and SiO2 was added. The resulting suspension was passed through a pad of Celite. The organic solvents were removed under reduced pressure to give a crude material, which was purified by preparative TLC (hexanes-EtOAc = 3:2) to afford diester 14 (2.01 mg, 2.30 μ mol, 42%). $R_f = 0.69$ (hexanes-EtOAc = 1:1). The physical data of diastereomeric mixture 14 is following: $[\alpha]_D^{29}$ -308 (c 0.201, CHCl₃); IR (neat, cm⁻¹): 2942, 2876, 1732, 1713, 1652, 1456, 1358, 1200, 1133, 1026, 998, 757; ¹H NMR (600 MHz, CDCl₃): δ 6.60 (dd, J = 11.4, 1.8 Hz, 2H), 6.31 (dt, J = 8.4, 2.4 Hz, 1H), 6.27 (dt, J = 8.4, 2.4 Hz, 1H), 5.74-5.62 (m, 2H), 5.09-5.04 (m, 2H), 4.89 (dd, J = 7.8, 1.8 Hz, 1H), 4.63-4.54 (m, 3H), 4.13-4.03(m, 2H), 3.94-3.77 (m, 4H), 3.52-3.30 (m, 3H), 3.02-2.84 (m, 1.5H), 2.77-2.56 (m, 1.5H), 1.84-1.70 (m, 2H), 1.70-1.44 (m, 14H), 1.20–1.10 (m, 2H), 1.15 (d, J = 7.2 Hz, 3H,), 1.09 (d, J = 6.6Hz, 3H), 0.97-0.85 (m, 12H); 13 C NMR (150 MHz, CDCl₂): δ 175.0, 174.0, 162.4, 141.2, 140.63, 140.62, 140.59, 139.2, 139.0, 105.73, 105.70, 105.2, 100.3, 99.0, 83.8, 81.1, 75.8, 69.7, 63.2, 63.15, 63.12, 63.02, 62.99, 62.4, 41.3, 41.1, 38.4, 38.2, 34.1, 34.0, 31.0, 30.9, 25.5, 25.10, 25.08, 20.1, 19.6, 15.6, 15.3, 11.8, 11.6, 10.7, 9.9, 0.0; HRMS (ESI) m/z: calcd for $C_{44}H_{60}N_2O_{12}S_2Na$ 895.3480 [M + Na⁺], found 895.3437.

(-)-Bis- β -hydroxy Ester 15. A screw cap test tube was charged with diester 14 (2.01 mg, 2.30 μ mol), MeOH (50 μ L), and CH₂Cl₂ (50 μ L). To the solution was added PPTS (2.31 mg, 9.20 μ mol), and the reaction mixture was stirred at room temperature for 5 h. The reaction mixture was purified by preparative TLC (hexanes-EtOAc = 3:2) to afford bis- β -hydroxy ester 15 (1.07 mg, 1.52 μ mol, 66%). $R_f = 0.32$ (hexanes-EtOAc = 1:1); $[\alpha]_D^{2\dot{5}}$ -254 (c 0.107, CHCl₃); IR (neat, cm⁻¹): 3253, 2963, 2934, 2877, 1709, 1653, 1362, 1140, 968, 755; ¹H NMR (600 MHz, CDCl₃): δ 6.61 (d, J = 1.9 Hz, 2H), 6.29 (dd, J = 1.8.2, 2.1 Hz, 2H), 5.62 (d, J = 8.6 Hz, 2H), 5.12 (dd, J = 8.6, 1.2 Hz, 2H), 4.54 (dd, J = 8.2, 1.2 Hz, 2H), 4.08 (d, J = 18.2 Hz, 2H), 3.65(dd, J = 9.1, 2.4 Hz, 2H), 2.93 (d, J = 18.2 Hz, 2H), 2.63 (qd, J = 7.2,2.4 Hz, 2H), 1.77 (dqd, J = 15.1, 7.4, 2.9 Hz, 2H), 1.48–1.40 (m, 2H), 1.20-1.14 (m, 2H), 1.12 (d, J = 7.2 Hz, 6H), 0.90 (t, J = 7.4 Hz, 6H), 0.81 (d, J = 6.7 Hz, 6H); ¹³C NMR (150 MHz, CDCl₃): δ 175.4, $162.6,\,141.3,\,139.5,\,113.3,\,105.3,\,75.7,\,74.7,\,70.3,\,62.7,\,41.9,\,37.1,\,34.0,\\$ 25.1, 15.0, 10.9, 9.1; HRMS (ESI) m/z: calcd for $C_{34}H_{44}N_2O_{10}S_2Na$ 727.2330 [M + Na⁺], found 727.2299.

(-)-Diester 16. A screw cap test tube was charged with (-)-deacetylaranotin (5) (2.06 mg, 4.90 μ mol), THP-protected β hydroxy carboxylic acid (+)-13 (5.01 mg, 20.5 μ mol), and dry CH₂Cl₂ (100 μ L). To the solution were added DMAP (1.85 mg, 15.1 μ mol) and WSCD·HCl (3.87 mg, 20.2 µmol). The reaction mixture was stirred at room temperature for 10 h. The reaction mixture was diluted with hexane and EtOAc, and SiO2 was added. The resulting suspension was passed through a pad of Celite. The organic solvents were removed under reduced pressure to give a crude material, which was purified by preparative TLC (hexanes-EtOAc = 1:1) to afford diester 16 (1.63 mg, 1.87 μ mol, 38%). $R_f = 0.61$ (hexanes-EtOAc = 1:1). The physical data of diastereomeric mixture (-)-16 is following: $[\alpha]_D^{24}$ -165 (c 0.0815, CHCl₃); IR (neat, cm⁻¹): 2939, 1731, 1715, 1354, 1141, 1026, 669; ¹H NMR (600 MHz, CDCl₃): δ 6.68–6.58 (m, 2H), 6.35-6.25 (m, 2H), 5.60 (d, J = 8.4 Hz, 1H), 5.55 (d, J = 9.0 Hz, 1H), 5.17-5.05 (m, 2H), 4.72 (d, J = 7.8 Hz, 1H,), 4.61-4.59 (m,

1H), 4.55 (d, J = 7.8 Hz, 1H), 4.54–4.49 (m, 1H), 4.10 (d, J = 18.0 Hz, 2H), 3.98–3.80 (m, 4H), 3.50–3.34 (m, 2.5H), 2.92 (d, J = 18.0 Hz, 2H), 2.72–2.56 (m, 1.5H), 1.86–1.75 (m, 2H), 1.75–1.42 (m, 14H), 1.32–1.20 (m, 2H), 1.20–1.14 (m, 6H), 0.98–0.82 (m, 12H); 13 C NMR (150 MHz, CDCl₃): δ 175.1, 170.2, 162.5, 162.4, 141.2, 140.8, 139.4, 139.2, 113.7, 105.7, 105.3, 104.9, 102.5, 101.2, 100.2, 82.7, 81.2, 75.7, 71.4, 70.3, 70.0, 63.5, 63.0, 62.7, 41.61, 41.55, 38.9, 37.6, 34.4, 34.0, 31.2, 31.1, 29.0, 25.5, 25.4, 25.1, 25.0, 20.8, 20.4, 15.7, 11.9, 11.8, 11.6, 11.0; HRMS (ESI) m/z: calcd for $C_{44}H_{60}N_2O_{12}S_2Na$ 895.3480 [M + Na⁺], found 895.3450.

(-)-SCH 64874 (17). A screw cap test tube was charged with diester 16 (1.63 mg, 1.87 μ mol), MeOH (50 μ L), and CH₂Cl₂ (50 μ L). To the solution was added PPTS (1.88 mg, 7.47 $\mu mol)\text{,}$ and the reaction mixture was stirred at 40 °C for 5 h. The reaction mixture was purified by preparative TLC (hexanes-EtOAc = 3:2) to afford 17 (1.14 mg, 1.62 μ mol, 86%). $R_f = 0.48$ (hexanes-EtOAc = 1:1); $[\alpha]_D^{29} - 169$ (c 0.141, CHCl₃); IR (neat, cm⁻¹): 3518, 2963, 2917, 1715, 1651, 1456, 1362, 1179, 1142, 969, 755; ¹H NMR (600 MHz, CDCl₃): δ 6.61 (d, J = 1.9 Hz, 2H), 6.30 (dd, J = 8.2, 2.1 Hz, 2H), 5.68 (ddd, J = 8.6, 2.1, 2.1 Hz, 2H), 5.09 (dd, I = 8.6, 1.9 Hz, 2H), 4.53 (dd, I = 8.2, 2.1 Hz, 2H), 4.08 (d, J = 18.2 Hz, 2H), 3.68 (d, J = 9.5 Hz, 2H), 2.92 (2H, dt, J = 18.2, 1.9 Hz), 2.63 (2H, qd, J = 7.2, 1.7 Hz), 1.81 (dqd, J = 15.3, 7.4, 3.1 Hz, 2H), 1.49–1.43 (m, 2H), 1.20–1.10 (m, 2H), 1.14 (d, J =7.8 Hz, 6H), 0.90 (t, J = 7.4 Hz, 6H), 0.79 (d, J = 6.7 Hz, 6H); 13 C NMR (150 MHz, CDCl₃): δ 175.5, 162.7, 141.4, 139.6, 113.1, 105.1, 75.7, 73.7, 69.9, 62.9, 41.6, 36.5, 33.9, 25.5, 14.9, 10.9, 8.8; HRMS (ESI) m/z: calcd for $C_{34}H_{44}N_2O_{10}S_2Na$ 727.2330 [M + Na⁺], found

(4S,6S)-β-Keto Ester 19. A screw cap test tube was charged with oxazolidinone 24¹⁸ (20 mg, 66 μ mol), dry MeOH (26 μ L, 0.66 μ mol), and dry THF (330 μ L). To the solution was added LiBH₄ (14 mg, 0.66 mmol) at 0 °C. The reaction mixture was warmed at room temperature and was stirred for 1 h. The reaction was quenched with 1 M NaOH aq. at 0 °C. The reaction mixture was diluted with water and extracted with EtOAc three times. The combined organic extracts were dried over Na₂SO₄, filtered, and concentrated under reduced pressure at 0 °C to give a crude material, which was purified by flash column chromatography (hexanes-EtOAc = 3:1) to afford alcohol 25 as a hexanes-EtOAc solution. All the solvent was not removed due to the volatility of the product. $R_f = 0.22$ (hexanes-EtOAc = 5:1). A screw cap test tube was charged with the hexanes-EtOAc solution of 25, Et₃N (46 μ L, 0.33 mmol), dry DMSO (110 μ L), and dry CH₂Cl₂ (220 μ L). To the solution was added SO₃·Py (52 mg, 0.33 mmol) at 0 °C. The resulting solution was stirred at 0 °C for 30 min. The reaction was quenched with water at 0 °C and then warmed to room temperature. The reaction mixture was extracted with CH2Cl2 three times. The combined organic extracts were dried over Na2SO4, filtered, and concentrated under reduced pressure at 0 °C to give a crude material as a CH2Cl2 solution, which was purified by flash column chromatography (hexanes-EtOAc = 5:1) to afford aldehyde 26 as a hexanes-EtOAc solution. All the solvent was not removed due to the volatility of the product. $R_f = 0.42$ (hexanes-EtOAc = 5:1). A screw-top test tube was charged with SnCl₂ (13 mg, 66 µmol) and dry CH₂Cl₂ (100 μ L). To the solution was added diazo ester 27 (17 mg, 0.13 mmol) and a hexanes-EtOAc solution of 26. After stirring at room temperature for 1.5 h, the reaction mixture was diluted with water and extracted with CH₂Cl₂ three times. The combined organic extracts were washed with water three times, dried over Na₂SO₄, filtered, and concentrated under reduced pressure to give a crude material, which was purified by flash column chromatography (hexanes-EtOAc = 5:1) to afford β -keto ester 19a and 19b (3.6 mg, 16 μ mol, 24% over 3 steps from 24) as a colorless oil. $R_f = 0.48$ (hexanes-EtOAc = 5:1). The physical data of a 1:1 mixture of 19a and 19b: IR (neat, cm⁻¹): 2963, 2930, 2876, 2855, 1747, 1738, 1732, 1715, 1462, 1456, 1377, 1246, 1194, 1121, 997; ¹H NMR (400 MHz, CDCl₃): δ 4.22–4.10 (m, 2H), 3.68 (q, J = 7.2 Hz, 0.5H), 3.63 (q, J = 7.2 Hz, 0.5H), 2.92–2.81 (m, 1H), 1.79 (ddd, J = 13.6, 8.4, 5.6 Hz, 0.5H), 1.68 (ddd, J = 13.6, 7.6, 6.0 Hz, 0.5H), 1.39–1.23 (m, 2.5H), 1.33 (d, *J* = 6.8 Hz, 1.5H), 1.32 (d, J = 7.2 Hz, 1.5H), 1.261 (dd, J = 7.2, 7.2 Hz, 1.5H), 1.257 (dd, J =7.2, 6.8 Hz, 1.5H), 1.16–1.00 (m, 2H), 1.10 (d, J = 6.8 Hz, 1.5H), 1.09 (d, J=6.8 Hz, 1.5H), 0.91–0.82 (m, 6.5H); ¹³C NMR (100 MHz, CDCl₃): δ 209.6, 170.5, 61.3, 61.2, 51.9, 51.3, 43.4, 43.3, 40.5, 39.7, 32.1, 32.0, 29.4, 29.2, 19.4, 19.3, 17.6, 17.1, 14.07, 14.05, 13.1, 12.9, 11.1, 11.0 (two signals are missing due to overlap); LRMS (EI) 228; HRMS (EI) m/z: calcd for $C_{13}H_{25}O_3$ 229.1804 [M + H⁺], found 229.1802.

(-)-(2S,4R)-Oxazolidinone 29. A flame-dried 50 mL two-necked round-bottomed flask was charged with dry THF (10 mL) and then NaHMDS (1.9 M in THF, 3.7 mL, 7.0 mmol) at -78 °C. To the solution was added oxazolidinone 28^{19} (1.45 g, 5.00 mmol) in THF (5 mL) via cannula at -78 °C over 5 min. The resulting solution was stirred for 1 h. To the solution was added MeI (1.56 mL, 25.0 mmol) dropwise at -78 °C over 3 min. After stirring at -78 °C for 1 h, the reaction was quenched with AcOH at −78 °C and then warmed to room temperature. The reaction mixture was diluted with water and extracted with EtOAc three times. The combined organic extracts were washed with water and brine, dried over Na₂SO₄, filtered, and concentrated under reduced pressure to give a crude material, which was purified by flash column chromatography (hexanes-EtOAc = 5:1) to afford oxazolidinone 29 (1.07 g, 3.53 mmol, 71%) as a colorless oil. $R_f = 0.22$ (hexanes-EtOAc = 5:1); $[\alpha]_D^{25}$ -55 (c 0.97, CHCl₃); IR (neat, cm⁻¹): 2962, 2929, 2874, 1782, 1698, 1455, 1386, 1350, 1238, 1208, 1099, 1015, 971, 760, 743, 702; 1 H NMR (600 MHz, CDCl₃): δ 7.33 (dd, J = 7.8, 7.2 Hz, 2H), 7.29–7.26 (m, 1H), 7.22 (d, J = 7.8 Hz, 2H), 4.67 (ddt, J = 9.6, 7.8, 2.4 Hz, 1H), 4.19 (dd, J = 8.4, 7.8 Hz, 1H), $4.17 \text{ (dd, } J = 9.0, 3.0 \text{ Hz, 1H)}, 3.82 \text{ (dt, } J = 9.0, 7.2 \text{ Hz, 1H)}, 3.27 \text{ (dd, } J = 9.0, 7.2 \text{ Hz, 1H})}$ I = 13.8, 2.4 Hz, 1H), 2.77 (dd, I = 13.2, 9.6 Hz, 1H), 1.58–1.51 (m, 1H), 1.45-1.29 (m, 3H), 1.23-1.13 (m, 1H), 1.20 (d, J = 6.6 Hz, 3H), 0.88 (d, J = 6.0 Hz, 3H), 0.87 (dd, J = 7.8, 6.6 Hz, 3H); 13 C NMR (150 MHz, CDCl₃): δ 177.7, 153.0, 135.3, 129.4, 128.9, 127.3, 66.0, 55.4, 40.1, 37.9, 35.4, 32.0, 29.8, 18.8, 17.2, 11.3; LRMS (EI) m/ z: 303 [M⁺]; HRMS (EI) m/z: calcd for $C_{18}H_{25}NO_3$ 303.1834 [M⁺], found 303.1819.

(4R,6S)-β-Keto Ester 20a and 20b. A screw cap test tube was charged with oxazolidinone 29 (20 mg, 66 μ mol), dry MeOH (26 μ L, 0.66 mmol), and dry THF (330 μ L). To the solution was added LiBH₄ (14 mg, 0.66 mmol) at 0 $^{\circ}\text{C}.$ After stirring at room temperature for 1 h, the reaction was quenched with 1 M NaOH aq. at 0 °C then warmed to room temperature. The mixture was diluted with water and extracted with EtOAc three times. The combined organic extracts were dried over Na2SO4, filtered, and concentrated under reduced pressure at 0 °C to give a crude material, which was purified by flash column chromatography (hexanes-EtOAc = 3:1) to afford alcohol 30 as a hexanes-EtOAc solution. All the solvent was not removed due to the volatility of the product. $R_f = 0.22$ (hexanes-EtOAc = 5:1). A screw cap test tube was charged with the hexanes-EtOAc solution of 30, Et₃N (46 μ L, 0.33 mmol), dry DMSO (110 μ L), and dry CH₂Cl₂ (220 μ L). To the solution was added SO₃·Py (52 mg, 0.33 mmol) at 0 °C. After stirring at 0 $^{\circ}\text{C}$ for 30 min, the reaction was quenched with water at 0 °C and then warmed to room temperature. The reaction mixture was extracted with CH2Cl2 three times. The combined organic extracts were dried over Na2SO4, filtered, and concentrated under reduced pressure at 0 °C to give a crude material in CH₂Cl₂, which was purified by flash column chromatography (hexanes-EtOAc = 5:1) to afford a crude aldehyde in hexanes-EtOAc solution. All the solvent was not removed due to the volatility of the product. $R_f = 0.42$ (hexanes-EtOAc = 5:1). A screw cap test tube was charged with SnCl₂ (13 mg, 66 μ mol) and dry CH₂Cl₂ (100 μ L), to which was added diazo ester 27 (17 mg, 0.13 mmol) and the crude aldehyde as a hexanes-EtOAc solution. After stirring at room temperature for 1.5 h, the reaction mixture was diluted with water and extracted with CH2Cl2 three times. The combined organic extracts were washed with water three times, dried over Na2SO4, filtered, and concentrated under reduced pressure to give a crude material, which was purified by flash column chromatography (hexanes-EtOAc = 5:1) to afford a mixture of β keto ester 20a and 20b (3.0 mg, 13 μ mol, 20% over 3 steps from 29) as a colorless oil. $R_f = 0.48$ (hexanes-EtOAc = 5:1). The physical data of a 1:1 mixture of **20a** and **20b**: IR (neat, cm⁻¹): 2964, 2936, 2876, 1747, 1715, 1462, 1456, 1377, 1244, 1194, 1120; ¹H NMR (400 MHz, CDCl₃): δ 4.18 (q, J = 7.2 Hz, 1H), 4.17 (q, J = 7.2 Hz, 1H), 3.67 (q, J

= 3.2 Hz, 0.5H), 3.65 (q, J = 7.2 Hz, 0.5H), 2.82 (dq, J = 12.8, 6.8 Hz, 1H), 1.53–1.43 (m, 0.5H), 1.43–1.24 (m, 2.5H), 1.33 (d, J = 7.2 Hz, 1.5H), 1.32 (d, J = 7.2 Hz, 1.5H), 1.26 (dd, J = 7.2, 7.2 Hz, 3H), 1.16 (dq, J = 14.4, 6.8 Hz, 0.5H), 1.12–1.05 (m, 1H), 1.07 (d, J = 6.8 Hz, 1.5H), 1.07 (d, J = 7.2 Hz, 1.5H), 0.91–0.82 (m, 6.5H); ¹³C NMR (100 MHz, CDCl₃): δ 209.9, 209.8, 170.6, 170.5, 61.3, 51.5, 51.3, 43.3, 43.2, 39.9, 39.2, 32.0, 31.8, 30.0, 29.8, 18.9, 18.8, 16.5, 15.9, 14.1, 13.1, 13.0, 11.31, 11.27 (two signals are missing due to overlap); LRMS (EI) 228; HRMS (EI) m/z: calcd for $C_{13}H_{25}O_3$ 229.1804 [M + H⁺], found 229.1801.

(+)-(25,3R,45,6S)-Oxazolidinone (32). A flame-dried 20 mL twonecked round-bottomed flask was charged with (3S)-3-propionyl-4benzyloxazolidin-2-one (296 mg, 1.27 mmol) and dry CH₂Cl₂ (1.2 mL). To the solution was added n-Bu₂BOTf (411 mg, 1.50 mmol) at 0 °C, and the resulting solution was stirred at 0 °C for 2 min. To the solution was added Et₂N (209 µL, 1.50 mmol) at 0 °C, and the solution was stirred at 0 $^{\circ}\text{C}$ for 10 min. To the solution was added a hexanes-EtOAc solution of aldehyde 26 dropwise at -78 °C, and the solution was stirred at -78 °C for 1 h and then at 0 °C for 1 h. The reaction was quenched with pH 7.0 phosphorus buffer (1 mL) and MeOH (4.1 mL), then MeOH-34% $\bar{H_2}O_2$ aq. (2:1, 4.1 mL) at 0 °C. The resulting solution was stirred at 0 °C for 1 h, then warmed to room temperature. The reaction mixture was diluted with water and extracted with Et₂O three times. The combined organic extracts were washed with 5% NaHCO3 and brine, dried over Na2SO4, filtered, and concentrated under reduced pressure at 0 °C to give a crude material, which was purified by flash column chromatography (hexanes-EtOAc = 5:1) to afford aldol product 32 (166 mg, 679 μ mol) as a white solid. In addition, the remaining alcohol 25 gave product 32 (380 mg, 994 μ mol). In total 546 mg (65%) of 32 was obtained from 24 over 3 steps. $R_f = 0.18$ (hexanes-EtOAc = 5:1); m.p.: 61–62 °C (hexanes, white prism); $[\alpha]_D^{26}$ +32.3 (c 1.57, CHCl₃); IR (neat, cm⁻¹): 3503, 2961, 2926, 2876, 1782, 1697, 1454, 1385, 1352, 1209, 1103, 970, 762, 748, 702; ¹H NMR (600 MHz, CDCl₃): δ 7.34 (t, J = 7.2 Hz, 2H), 7.28 (t, J = 7.2 Hz, 1H), 7.21 (d, J = 7.2 Hz, 2H), 4.72–4.67 (m, 1H), 4.23 (dd, J = 9.0, 7.2 Hz, 1H), 4.19 (dd, J = 9.0, 3.0 Hz, 1H), 3.99 (ddd, J = 14.4, 7.2, 4.8 Hz, 1H), 3.67 (dd, J = 6.0, 4.2 Hz, 1H), 3.26 (dd, J = 13.2, 3.0 Hz, 1H), 2.79 (dd, J = 13.2, 3.0 Hz, 1H), 2.46 (br s, 1H), 1.70-1.63 (m, 1H), 1.51-1.46 (m, 1H), 1.45-1.38 (m, 1H), 1.32 (ddd, J = 13.2, 7.2, 4.8 Hz, 1H), 1.28 (d, J = 7.8 Hz, 3H), 1.06 (dq, J = 15.6, 7.8 Hz, 1H), 0.98-0.92 (m, 1H), 0.96 (d, J = 6.0 Hz,3H), 0.88 (d, J = 6.0 Hz, 3H), 0.88–0.86 (m, 3H); ¹³C NMR (150 MHz, CDCl₃): δ 177.4, 152.9, 135.0, 129.4, 128.9, 127.4, 75.2, 66.1, 55.1, 40.4, 40.0, 37.7, 33.1, 31.4, 28.3, 20.0, 15.4, 11.7, 11.0; LRMS (EI) 361.2 [M⁺]; HRMS (EI) m/z: calcd for $C_{21}H_{31}NO_4$ 361.2253 [M⁺], found 361.2243.

(-)-(2S,3R,4S,6S)- β -Hydroxy Carboxylic Acid **33**. A screw cap test tube was charged with H_2O_2 (34% w/w solution in H_2O , 39 μ L, 0.39 mmol), LiOH·H₂O (3.5 mg, 83 μ mol), and H₂O (45 μ L). To the solution was added aldol product 32 (20.0 mg, 55.3 μ mol) in THF (164 μ L) dropwise at 0 °C. After stirring at room temperature for 15 min, the reaction was quenched with saturated aqueous Na₂S₂O₃ and 1 M aqueous HCl. The aqueous layer was extracted with EtOAc three times. The combined organic extracts were dried over Na₂SO₄, filtered, and concentrated under reduced pressure to give a crude material, which was purified by preparative TLC (CH₂Cl₂-EtOAc = 20:1) to afford β -hydroxy carboxylic acid 33 (10.0 mg, 47.5 μ mol, 86%) as a colorless oil. $R_f = 0.32$ (hexanes-EtOAc = 1:1); $[\alpha]_D^{27} - 16.8$ (c 1.24, CHCl₃); IR (neat, cm⁻¹): 3403, 2962, 2921, 2877, 2852, 1710, 1460, 1380, 1204, 982; ¹H NMR (600 MHz, CDCl₃): δ 3.69 (dd, J = 5.4, 5.4 Hz, 1H), 2.73 (dq, J = 7.2, 5.4 Hz, 1H), 1.73-1.65 (m, 1H), 1.50-1.43 (m, 1H), 1.43–1.35 (m, 1H), 1.32–1.23 (m, 1H), 1.25 (d, *J* = 7.8 Hz, 3H), 1.10-1.01 (m, 1H), 0.99 (m, 1H), 0.95 (d, J = 6.0 Hz, 3H), 0.89–0.84 (m, 6H); 13 C NMR (150 MHz, CDCl₃): δ 181.5, 75.3, 42.4, 40.6, 33.0, 31.2, 28.5, 19.8, 14.7, 11.7, 11.0; LRMS (FAB⁺) 203; HRMS (FAB+) m/z: calcd for $C_{11}H_{23}O_3$ 203.1669 [M + H+], found

(–)-(2S,3R,4S,6S)-Ethyl Ester **34**. A screw cap test tube was charged with β -hydroxy carboxylic acid **33** (5.0 mg, 25 μ mol) and acetone (411 μ L). To the solution was added EtI (25.0 μ L, 247 μ mol) and K₂CO₃

(17.0 mg, 124 μ mol) at room temperature. After stirring at 50 °C for 3 h, the reaction mixture was diluted with CHCl₂ and filtered through a pad of Celite. The filter cake was washed with CHCl3. The filtrate was concentrated under reduced pressure to give a crude material, which was purified by preparative TLC (EtOAc) to afford ethyl ester 34 (5.0 mg, 22 μ mol, 89%) as a colorless oil. $R_f = 0.73$ (hexanes-EtOAc = 1:1); $[\alpha]_{\rm D}^{27}$ –12 (c 0.90, CHCl₃); IR (neat, cm⁻¹): 3507, 2963, 2930, 2877, 2855, 1733, 1715, 1462, 1376, 1336, 1254, 1181, 1162, 1114, 1095, 1046, 985; 1 H NMR (600 MHz, CDCl $_{3}$): δ 4.19–4.10 (m, 2H), 3.62 (br s, 1H), 2.65 (dq, J = 6.6, 6.6 Hz, 1H), 2.09 (br s, 1H), 1.61 (dq, J =12.6, 6.6 Hz, 1H), 1.49–1.34 (m, 2H), 1.31–1.22 (m, 1H), 1.27 (dd, J = 8.4, 6.0 Hz, 3H), 1.21 (d, J = 6.6 Hz, 3H), 1.10-1.00 (m, 1H),0.98-0.88 (m, 1H), 0.93 (d, J = 6.6 Hz, 3H), 0.88-0.80 (m, 6H); 13 C NMR (150 MHz, CDCl₃): δ 176.1, 75.3, 60.5, 42.6, 40.7, 32.9, 31.2, 28.5, 19.8, 14.5, 14.2, 12.2, 11.0; LRMS (FAB+) 231; HRMS (FAB+) m/z: calcd for C₁₃H₂₇O₃ 231.1960 [M + H⁺], found 231.1963.

(–)-(2S,3R,4S,6S)-THP-Protected β -Hydroxy Carboxylic Acid **41**. A flame-dried 50 mL two-necked round-bottomed flask was charged with oxazolidinone 32 (180 mg, 498 μ mol), DHP (70.0 μ L, 767 μ mol), and dry CH₂Cl₂ (3.3 mL). To the solution was added PPTS (12.5 mg, 48.9 μ mol). After stirring at room temperature for 20 min, additional DHP (70.0 μ L, 767 mmol) and PPTS (12.5 mg, 48.9 μ mol) were added. After stirring for additional 15 min, PPTS (12.5 mg, 48.9 μ mol) was added, and the stirring was continued for more 3 h. The reaction mixture was diluted with CH2Cl2. The separated organic layer was washed twice with 13.2% aqueous sodium chloride, dried over Na₂SO₄, filtered, and concentrated under reduced pressure to give a crude material as a colorless oil, which was used for the next reaction without further purification. $R_f = 0.47$ (hexanes-EtOAc = 3:1). A 50 mL two-necked round-bottomed flask was charged with the crude material, THF (6.75 mL), and H₂O (2.25 mL). To the mixture was added LiOH·H₂O (41 mg, 1.0 mmol) and H₂O₂ (34% w/w solution in H₂O, 0.29 mL, 2.9 mmol) at 0 °C and warmed at room temperature for 1 h. Additional LiOH·H₂O (41 mg, 1.0 mmol) and H₂O₂ (34% w/ w solution in H_2O , 291 μL , 2.91 mmol) were added at 0 °C. After stirring at room temperature for 6 h, the reaction was quenched with saturated aqueous Na₂S₂O₃ and 1 M aqueous HCl. The separated aqueous layer was extracted with EtOAc three times. The combined organic extracts were dried over Na2SO4, filtered, and concentrated under reduced pressure to give a crude material, which was purified by flash column chromatography (hexanes-EtOAc = 3:1) to afford THPprotected β -hydroxy carboxylic acid 41 (127 mg, 443 μ mol, 89%) as a colorless oil. $R_f = 0.12$ (hexanes-EtOAc = 3:1); $[\alpha]_D^{27} + 12.3$ (c 3.04, CHCl₃); IR (neat, cm⁻¹): 2957, 2876, 2853, 1711, 1705, 1462, 1456, 1132, 1034, 1026, 997; ¹H NMR (600 MHz, CDCl₃): δ 4.63 (dd, J = 7.2, 3.0 Hz, 0.5H), 4.55 (dd, J = 4.8, 2.4 Hz, 0.5H), 4.00–3.93 (m, 1H), 3.82 (dd, J = 6.0, 3.6 Hz, 0.5H), 3.74 (dd, J = 5.4, 3.6 Hz, 0.5H), 3.53-3.48 (m, 1H), 2.93 (dq, J = 13.2, 6.6 Hz, 0.5H), 2.76 (dq, J = 13.2) 13.2, 6.6 Hz, 0.5H), 1.87-1.71 (m, 3H), 1.62-1.45 (m, 7.5H), 1.40-1.31 (m, 1.5H), 1.20 (d, I = 7.2 Hz, 1.5H), 1.19 (d, I = 7.8 Hz, 1.5H), 1.15-1.05 (m, 0.5H), 1.02-0.94 (m, 0.5H), 0.94 (d, J = 6.6 Hz, 1.5H), 0.89–0.84 (m, 7.5H); 13 C NMR (150 MHz, CDCl₃): δ 181.5, 180.1, 101.8, 100.4, 82.9, 81.7, 64.2, 63.4, 42.7, 42.4, 41.1, 40.9, 33.9, 33.2, 31.5, 31.4, 31.1, 30.9, 29.1, 28.7, 25.3, 25.1, 20.8, 20.2, 19.8, 19.4, 15.4, 15.1, 13.1, 12.6, 11.1, 11.0; LRMS (FAB+) 287; HRMS (FAB+) m/z: calcd for $C_{16}H_{30}O_4$ 287.2222 [M + H⁺], found 287.2218.

(*–*)-*Diol* **35**. A screw cap test tube was charged with diacetate **36** (12.0 mg, 22.4 μmol) and MeOH (224 μL). To the solution was added K_2CO_3 (15.0 mg, 112 μmol). After stirring at room temperature for 0.5 h, the reaction mixture was diluted with water and extracted with EtOAc three times. The combined organic extracts were dried over Na_2SO_4 , filtered, and concentrated under reduced pressure to give a crude material, which was purified by preparative TLC (CH₂Cl₂-acetone =20:1) to afford diol **35** (10.1 mg, 22.4 μmol, quant.) as a white solid. R_f = 0.36 (CH₂Cl₂-acetone = 10:1); $[\alpha]_D^{27}$ –274 (c 0.305, CHCl₃); IR (neat, cm⁻¹): 3393, 2918, 1691, 1661, 1651, 1404, 1339, 1196, 1130, 910, 731; ¹H NMR (600 MHz, CDCl₃): δ 6.55 (s, 2H), 6.23 (dd, J = 7.8, 1.8 Hz, 2H), 4.92 (dd, J = 7.8, 1.8 Hz, 2H), 4.88–4.84 (m, 2H), 4.68 (d, J = 7.8 Hz, 2H), 4.59 (br s, 2H), 3.04 (s, 4H), 2.28 (s, 6H); ¹³C NMR (150 MHz, CDCl₃): δ 166.5, 138.1, 137.9,

110.8, 107.6, 72.6, 69.3, 64.1, 39.5, 14.8; HRMS (ESI⁺) m/z: calcd for $C_{20}H_{22}N_2O_6S_2Na$ 473.0817 [M + Na⁺], found 473.0796.

THP-Protected β -Hydroxy Ester 42. A screw cap test tube was charged with core skeleton 35 (8.7 mg, 19 μ mol), carboxylic acid 41 (5.5 mg, 19 μ mol), and CH₂Cl₂ (100 μ L). To the solution was added DMAP (2.3 mg, 19 μ mol) and WSCD·HCl (7.4 mg, 39 μ mol) at 0 °C. After stirring at reflux for 2 h, the reaction was quenched with 1 M HCl aq. The resulting mixture was extracted with CH₂Cl₂ three times. The combined organic extracts were washed with H₂O and brine, dried over Na₂SO₄, filtered, and concentrated under reduced pressure. The residue was purified by preparative TLC (CH₂Cl₂-acetone = 20:1) to afford THP-protected β -hydroxy ester 42 (5.0 mg, 7.0 μ mol, 36%) as a colorless oil with recovery of the core skeleton 35 (2.6 mg, 5.8 μ mol, 30%). $R_f = 0.47$ (CH₂Cl₂-acetone = 10:1); $[\alpha]_D^{27} - 224$ (c 0.640, CHCl₃); IR (neat, cm⁻¹): 3420, 2957, 2928, 1732, 1693, 1681, 1666, 1651, 1633, 1392, 1128, 1026, 756; ¹H NMR (500 MHz, CDCl₃): δ 6.60–6.55 (m, 1H), 6.52 (d, J = 1.5 Hz, 1H), 6.32–6.25 (m, 1H), 6.23-6.19 (m, 1H), 5.78 (ddd, J = 8.0, 2.0, 2.0 Hz 0.4H), 5.75 (ddd, I = 8.0, 2.0, 2.0 Hz, 0.6H), 5.23-5.13 (m, 1H), 4.95-4.91(m, 1H), 4.89 (d, J = 7.5 Hz, 1H), 4.70-4.55 (m, 2H), 4.51-4.40 (m, 1H)1H), 3.96-3.83 (m, 2H), 3.50-3.42 (m, 0.6H), 3.43-3.36 (m, 0.4H), 3.14-2.93 (m, 4H), 2.71-2.60 (m, 1H), 2.30-2.27 (m, 6H), 1.85-1.75 (m, 1H), 1.75–1.65 (m, 1H), 1.67–1.60 (m, 1H), 1.60–1.42 (m, 5H), 1.42–1.33 (m, 1H), 1.31 (d, J = 7.0 Hz, 1.2H), 1.25 (d, J = 7.5 Hz, 1.8H), 1.15 (dd, J = 7.0, 4.0 Hz, 1H), 1.13–0.90 (m, 2H), 0.95 (d, $J = 6.0 \text{ Hz}, 1.8 \text{H}), 0.92 \text{ (d, } J = 9.0 \text{ Hz}, 1.2 \text{H}), 0.89 - 0.80 \text{ (m, 6H)}; ^{13}\text{C}$ NMR (125 MHz, CDCl₃): δ 174.6, 174.3, 166.9, 164.10, 164.5, 139.6, 139.3, 137.94, 137.92, 137.8, 137.6, 110.8, 109.7, 109.6, 107.9, 107.8, 106.1, 105.8, 102.0, 100.5, 83.2, 81.7, 72.60, 72.57, 72.1, 71.8, 70.57, 70.54, 69.30, 69.26, 64.17, 64.15, 63.5, 60.6, 60.5, 60.3, 43.2, 42.6, 41.2, 41.1, 40.6, 40.5, 40.3, 40.2, 34.3, 34.1, 31.48, 31.47, 31.3, 31.1, 29.1, 28.8, 28.3, 25.4, 25.3, 21.1, 20.5, 20.2, 19.9, 19.8, 1.6, 15.5, 14.9, 14.83, 14.81, 14.79, 14.6, 12.7, 12.1, 11.2, 11.1, 11.0; HRMS (ESI⁺) m/z: calcd for $C_{36}H_{50}N_2O_9S_2Na$ 741.2855 [M + Na⁺], found 741.2820.

 β -Hydroxy Ester 43. A screw cap test tube was charged with 42 (11.0 mg, 15.3 μ mol), MeOH (76 μ L), and CH₂Cl₂ (76 μ L). To the solution was added PPTS (7.7 mg, 31 μ mol). After stirring at 40 °C for 1.5 h, the reaction mixture was diluted with CH2Cl2. The organic layer was washed twice with 13.2% aqueous sodium chloride, dried over Na₂SO₄, filtered, and concentrated under reduced pressure to give a crude material, which was purified by preparative TLC (CH₂Cl₂acetone = 20:1) to afford the desired β -hydroxy ester 43 (7.6 mg, 12 μ mol, 78%) as a pale yellow solid. $R_f = 0.49$ (CH₂Cl₂-acetone =10:1); $[\alpha]_{\rm D}^{21}$ -243 (c 0.760, CHCl₃); IR (neat, cm⁻¹): 3479, 2961, 2924, 2874, 1728, 1693, 1666, 1659, 1393, 1339, 1194, 1126, 756; ¹H NMR (500 MHz, CDCl₃): δ 6.61 (dd, J = 2.5, 2.0 Hz, 1H), 6.51 (s, 1H), 6.32 (dd, J = 8.5, 2.0 Hz, 1H), 6.21 (dd, J = 8.5, 2.5 Hz, 1H), 5.88 (ddd, J = 8.0, 2.0, 2.0 Hz, 1H), 5.16 (d, J = 7.5 Hz, 1H), 4.93 (dd, J = 7.5 Hz, 1H)8.0, 2.0 Hz, 1H), 4.91-4.87 (m, 1H), 4.71-4.66 (m, 1H), 4.62, (dd, J = 8.5, 2.0 Hz, 1H), 4.43 (br s, 1 H), 3.77 (dd, J = 8.0, 2.5 Hz, 1H),3.14-2.95 (m, 4H), 2.72 (dq, J = 7.5, 3.0 Hz, 1H), 2.31 (s, 3H), 2.30(s, 3H), 1.67-1.59 (m, 1H), 1.55-1.40 (m, 3H), 1.22 (d, J = 7.0 Hz,3H), 1.21-1.15 (m, 1H), 1.04-0.97 (m, 1H), 0.99 (d, J = 6.0 Hz, 3H), 0.94–0.82 (m, 7H); 13 C NMR (125 MHz, CDCl₃): δ 175.1, 166.8, 164.8, 139.9, 138.1, 138.0, 137.6, 110.8, 109.4, 107.8, 105.6, 74.7, 72.5, 71.8, 70.3, 69.6, 64.2, 60.8, 42.3, 40.6, 40.5, 40.4, 33.1, 31.3, 27.7, 20.3, 15.9, 14.9, 14.8, 10.9, 9.8; HRMS (ESI+) m/z: calcd for $C_{31}H_{42}N_2O_8S_2Na$ 657.2280 [M + Na⁺], found 657.2269.

(–)-(2R,4R)-Oxazolidinone 45. A flame-dried 50 mL two-necked round-bottomed flask was charged with dry THF (7 mL). To the solution was added NaHMDS (1.9 M in THF, 2.6 mL, 4.9 mmol) at -78 °C and then oxazolidinone 44 (1.02 g, 3.52 mmol) in THF (5 mL) via cannula at -78 °C over 5 min. After stirring stirred at -78 °C for 0.5 h, MeI (1.09 mL, 17.6 mmol) was added dropwise at -78 °C over 3 min. After stirring at -78 °C for 2 h, the reaction was quenched with AcOH at -78 °C and allowed to warm to room temperature. The reaction mixture was diluted with water and extracted with EtOAc three times. The combined organic extracts were washed with water and brine, dried over Na₂SO₄, filtered, and concentrated under reduced pressure to give a crude material, which was purified by flash

column chromatography (hexanes-EtOAc = 5:1) to afford oxazolidinone **45** (909 mg, 2.99 mmol, 85%) as a colorless oil. $R_f=0.28$ (hexanes-EtOAc = 5:1); $[\alpha]_D^{27}-68.6$ (c 4.13, CHCl₃); IR (neat, cm⁻¹): 2962, 2929, 2874, 1781, 1698, 1456, 1387, 1350, 1289, 1241, 1209, 1099, 1016, 973, 742, 702; ¹H NMR (600 MHz, CDCl₃): δ 7.34 (t, J=7.8 Hz, 2H), 7.29–7.24 (m, 1H), 7.22 (d, J=7.8 Hz, 2H), 4.69 (ddt, J=9.6, 7.2, 3.0 Hz, 1H), 4.20 (dd, J=9.0, 7.8 Hz, 1H), 4.17 (dd, J=9.0, 3.0 Hz, 1H), 3.90–3.84 (m, 1H), 3.26 (dd, J=13.2, 3.0 Hz, 1H), 1.42–1.29 (m, 2H), 1.22 (d, J=7.2 Hz, 3H), 1.19–1.09 (m, 2H), 0.88 (dd, J=7.8, 6.6 Hz, 3H), 0.86 (d, J=6.0 Hz, 3H); ¹³C NMR (150 MHz, CDCl₃): δ 177.3, 152.9, 135.2, 129.4, 128.8, 127.2, 65.9, 55.2, 40.4, 37.8, 35.3, 32.2, 29.3, 19.3, 18.3, 11.2; LRMS (EI) m/z: 303 [M⁺]; HRMS (EI) m/z: calcd for $C_{18}H_{25}NO_3$ 303.1834 [M⁺], found 303.1822. Spectral data matched the reported data.

(2R,3S,4R,6R)-Oxazolidinone 49. A flame-dried 30 mL two-necked round-bottomed flask was charged with oxazolidinone 45 (400 mg, 1.32 mmol), dry MeOH (266 μ L, 6.59 mmol), and dry THF (6.6 mL). To the solution was added LiBH₄ (143 mg, 6.59 mmol) at 0 °C. After stirring at room temperature for 5 h, the reaction was quenched with 1 M NaOH aq. at 0 °C then warmed to room temperature. The reaction mixture was diluted with water and extracted with EtOAc three times. The combined organic extracts were dried over Na₂SO₄, filtered, and concentrated under reduced pressure at 0 °C to give a crude material, which was purified by flash column chromatography (hexanes-EtOAc = 3:1) to afford alcohol 46 as a hexanes-EtOAc solution. All the solvent was not removed due to the volatility of the product. $R_f = 0.22$ (hexanes-EtOAc = 5:1). A flame-dried 20 mL two-necked roundbottomed flask was charged with a hexanes-EtOAc solution 46, Et₃N (918 μL, 6.59 mmol), dry DMSO (1.3 mL), and dry CH₂Cl₂ (2.6 mL). To the solution was added SO₃·Py (1.05 g, 6.59 mmol) at 0 °C. After stirring at 0 °C for 15 min, the reaction was quenched with water at 0 °C. The reaction mixture was extracted with CH₂Cl₂ three times. The combined organic extracts were dried over Na2SO4, filtered, and concentrated under reduced pressure at 0 °C to give a crude material, which was purified by flash column chromatography (hexanes-EtOAc = 3:1) to afford aldehyde 47 as a hexanes-EtOAc solution. All the solvent was not removed due to the volatility of the product. $R_f = 0.42$ (hexanes-EtOAc = 5:1). A flame-dried 20 mL two-necked roundbottomed flask was charged with (3R)-3-propionyl-4-benzyloxazolidin-2-one (338 mg, 1.45 mmol) and dry CH₂Cl₂ (1.3 mL). To the solution was added n-Bu₂BOTf (469 mg, 1.71 mmol) at 0 °C. After stirring at 0 °C for 10 min, Et₃N (239 μ L, 1.71 mmol) was added at 0 °C. Then, after stirring at 0 °C for 10 min, a hexanes-EtOAc solution of 48 was added dropwise at -78 °C. The resulting solution was stirred at -78 °C for 1 h and then warmed to 0 °C over 35 min. The reaction was quenched with pH 7.0 phosphorus buffer (1.5 mL) and MeOH (4 mL), then MeOH-34% H_2O_2 aq. (2:1, 6 mL) at 0 °C. The resulting solution was stirred at 0 °C for 1 h and then warmed to room temperature. The reaction mixture was diluted with water and extracted with Et₂O three times. The combined organic extracts were washed with 5% NaHCO3 and brine, dried over Na2SO4, filtered, and concentrated under reduced pressure at 0 °C to give a crude material, which was purified by flash column chromatography (hexanes-EtOAc = 5:1) to afford oxazolidinone 49 (213 mg, 593 μ mol, 45% over 3 steps from 45) as a white solid. $R_f = 0.18$ (hexanes-EtOAc = 5:1); m.p.: 62–63 °C (hexanes, white prism); $[\alpha]_D^{27}$ –34.5 (c 1.54, CHCl₃). Spectral data matched the compound 32.

(–)-(2R,3S,4R,6R)-Carboxylic Acid **50**. A flame-dried 20 mL two-necked round-bottomed flask was charged with oxazolidinone **49** (190 mg, 526 μ mol), DHP (144 μ L, 1.58 mmol), and dry CH₂Cl₂ (3.5 mL). To the solution was added PPTS (53 mg, 0.21 mmol). After stirring at room temperature for 1 h, additional DHP (48.0 μ L, 526 μ mol) and PPTS (13.3 mg, 52.6 μ mol) were added. After stirring at room temperature for 0.5 h, the reaction mixture was diluted with CH₂Cl₂. The separated organic layer was washed twice with 13.2% aqueous sodium chloride, dried over Na₂SO₄, filtered, and concentrated under reduced pressure to give a crude material as a colorless oil, which was used to the next reaction without further purification. $R_f = 0.47$ (hexanes-EtOAc = 3:1). A 20 mL two-necked round-bottomed flask

was charged the crude material, THF (2 mL), and H₂O (0.6 mL). To the solution was added H₂O₂ (34% w/w solution in H₂O, 268 μ L, 2.63 mmol) and LiOH·H₂O (44 mg, 1.1 mmol) at 0 °C. After stirring at room temperature for 10 h, the reaction was quenched with saturated aqueous Na₂S₂O₃ and 1 M HCl aq. The separated aqueous layer was extracted with EtOAc three times. The combined organic extracts were dried over Na₂SO₄, filtered, and concentrated under reduced pressure to give a crude material, which was purified by flash column chromatography (hexanes-EtOAc = 3:1) to afford carboxylic acid **50** (98.1 mg, 341 μ mol, 65% over 2 steps from **49**) as a as a colorless oil. $R_f = 0.12$ (hexanes-EtOAc = 3:1); $[\alpha]_D^{27}$ -12.7 (c 3.35, CHCl₃). Spectral data matched the compound **41**.

(-)-(2R,3S,4S,6S)-Oxazolidinone **S4** (for Preparation of β -Hydroxy Carboxylic Acid 54). A flame-dried 30 mL two-necked round-bottomed flask was charged with oxazolidinone 24 (400 mg, 1.32 mmol), dry MeOH (266 μ L, 6.59 mmol), and dry THF (6.6 mL). To the solution was added LiBH₄ (143 mg, 6.59 μ mol) at 0 °C. After stirring at room temperature for 5 h, the reaction was quenched with 1 M NaOH at 0 °C. The reaction mixture was diluted with water and extracted with EtOAc three times. The combined organic extracts were dried over Na2SO4, filtered, and concentrated under reduced pressure at 0 °C to give a crude material, which was purified by flash column chromatography (hexanes-EtOAc = 3:1) to afford alcohol 25 as a hexanes-EtOAc solution. All the solvent was not removed due to the volatility of the product. $R_f = 0.22$ (hexanes-EtOAc = 5:1). A flamedried 20 mL two-necked round-bottomed flask was charged with a hexanes-EtOAc solution of 25, Et₃N (918 µL, 6.59 mmol), dry DMSO (1.3 mL), and dry CH₂Cl₂ (2.6 mL). To the solution was added SO₃· Py (1.05 g, 6.59 mmol) at 0 °C. After stirring at 0 °C for 10 min, the reaction was quenched with water at 0 °C. The reaction mixture was extracted with CH2Cl2 three times. The combined organic extracts were dried over Na₂SO₄, filtered, and concentrated under reduced pressure at 0 °C to give a crude material, which was purified by flash column chromatography (hexanes-EtOAc = 3:1) to afford aldehyde 26 as a hexanes-EtOAc solution. All the solvent was not removed due to the volatility of the product. $R_f = 0.42$ (hexanes-EtOAc = 5:1). A flame-dried 20 mL two-necked round-bottomed flask was charged with (3R)-3-propionyl-4-benzyloxazolidin-2-one (338 mg, 1.45 mmol) and dry CH₂Cl₂ (1.3 mL). To the solution was added n-Bu₂BOTf (469 mg, 1.71 mmol) at 0 °C. After stirring at 0 °C for 10 min, Et₃N (239 μL ,1.71 mmol) was added at 0 °C, and the resulting solution was stirred at 0 °C for 10 min. Then, a hexanes-EtOAc solution 26 was added dropwise at -78 $^{\circ}$ C, and the resulting solution was stirred at -78 °C for 1 h and then 0 °C for 0.5 h. The reaction was guenched with pH 7.0 phosphorus buffer (1.5 mL) and MeOH (4 mL), then MeOH-34% H₂O₂ aq. (2:1, 6 mL) at 0 °C. The resulting solution was stirred at 0 °C for 1 h and then warmed to room temperature. The reaction mixture was diluted with water and extracted with Et₂O three times. The combined organic extracts were washed with 5% NaHCO₃ and brine, dried over Na2SO4, filtered, and concentrated under reduced pressure at 0 $^{\circ}\text{C}$ to give a crude material, which was purified by flash column chromatography (CH₂Cl₂) to afford S4 (390 mg, 1.08 mmol, 82% over 3 steps from 24) as a white solid. $R_f = 0.15$ (hexanes-EtOAc = 5:1); $[\alpha]_D^{27}$ -56.1 (c 2.04, CHCl₃); m.p.: 86–87 °C (hexanes, white prism); IR (neat, cm⁻¹): 3524, 2961, 2930, 2874, 1782, 1697, 1454, 1385, 1350, 1236, 1209, 1107, 982, 970, 762, 702; ¹H NMR (600 MHz, CDCl₃): δ 7.36–7.32 (m, 2H), 7.30–7.25 (m, 1H), 7.22– 7.19 (m, 2H), 4.69 (ddt, J = 9.6, 7.8, 3.6 Hz, 1H), 4.21 (dd, J = 9.0, 7.8 Hz, 1H), 4.19 (dd, J = 9.0, 3.0 Hz, 1H), 3.97 (dq, J = 7.2, 3.0 Hz, 1H), 3.59 (dd, J = 8.4, 3.0 Hz, 1H), 3.26 (dd, J = 13.2, 3.6 Hz, 1H), 2.97(dd, J = 13.2, 9.6 Hz, 1H), 2.72 (br s, 1H), 1.68-1.60 (m, 2H), 1.49-1.40 (m, 2H), 1.25 (d, J = 6.6 Hz, 3H), 1.06–0.98 (m, 1H), 0.96–0.91 (m, 1H), 0.90–0.85 (m, 9H); 13 C NMR (150 MHz, CDCl₃): δ 177.8, 152.8, 135.0, 129.4, 129.0, 127.4, 76.1, 66.1, 55.2, 40.2, 39.7, 37.8, 33.5, 31.8, 27.9, 20.4, 16.0, 11.1, 10.3; LRMS (EI) 361; HRMS (EI) *m/z*: calcd for $C_{21}H_{31}NO_4$ 361.2253 [M $^+$], found 361.2235.

(–)-(2R, 3S, 4S, 6S)-THP-Protected β -Hydroxy Carboxylic Acid **54**. A flame-dried 20 mL two-necked round-bottomed flask was charged with the above oxazolidinone **S4** (190 mg, 526μ mol), DHP (144 μ L, 1.58 mmol), and dry CH₂Cl₂ (3.5 mL). To the solution was added PPTS

(53 mg, 0.21 mmol). After stirring at room temperature for 1.5 h, the reaction was diluted with CH2Cl2. The separated organic layer was washed twice with 13.2% aqueous sodium chloride, dried over Na₂SO₄, filtered, and concentrated under reduced pressure to give a crude material as a colorless oil, which was used to the next reaction without further purification. $R_f = 0.33$ (hexanes-EtOAc = 5:1). A 20 mL two-necked round-bottomed flask was charged with the crude material, THF (2.0 mL), and H₂O (0.6 mL). To the solution was added H₂O₂ (34% w/w solution in H₂O, 0.27 mL, 2.6 mmol) and LiOH·H₂O (44 mg, 1.1 mmol) at 0 °C. After stirring at room temperature for 9 h, the reaction was quenched with saturated aqueous Na₂S₂O₃ and 1 M aqueous HCl. The separated aqueous layer was extracted with EtOAc three times. The combined organic extracts were dried over Na2SO4, filtered, and concentrated under reduced pressure to give a crude material, which was purified by flash column chromatography (hexanes-EtOAc = 3:1) to afford THP-protected β hydroxy carboxylic acid 54 (83.0 mg, 289 mmol, 55% over 2 steps from the oxazolidinone S4) as a colorless oil. $R_f = 0.19$ (hexanes-EtOAc = 3:1); $[\alpha]_D^{27}$ -24.3 (c 2.35, CHCl₃); IR (neat, cm⁻¹): 2957, 2875, 2854, 1737, 1708, 1462, 1381, 1282, 1234, 1201, 1167, 1131, 1077, 1084, 998, 962, 905, 869, 813; 1 H NMR (500 MHz, CDCl₃): δ 4.63 (dd, J = 5.0, 3.0 Hz, 0.5H), 4.56 (dd, J = 6.0, 2.0 Hz, 0.5H), 3.95-3.89 (m, 1H), 3.85 (dd, J = 5.0, 5.0 Hz, 0.5H), 3.74 (dd, J = 5.0, 5.0 Hz, 0.5H), 3.52-3.43 (m, 1H), 2.82 (dq, J = 7.0, 4.5 Hz, 0.5H), 2.71(dq, J = 7.0, 4.5 Hz, 0.5H), 1.89-1.69 (m, 3H), 1.61-1.33 (m, 7.5H),1.22 (d, J = 7.5 Hz, 1.5H), 1.19 (d, J = 7.0 Hz, 1.5H), 1.07-0.65 (m, 1.5H), 0.97 (d, I = 7.0 Hz, 1.5H), 0.91 (d, I = 7.0 Hz, 1.5H), 0.89– 0.83 (m, 6H); 13 C NMR (125 MHz, CDCl₃): δ 181.9, 180.6, 101.0, 100.2, 83.7, 82.1, 63.6, 63.1, 41.7, 41.3, 39.7, 39.5, 34.9, 33.5, 31.7, 31.6, 31.0, 30.9, 28.4, 28.0, 25.4, 25.2, 20.5, 20.2, 19.98, 19.97, 16.6, 16.5, 12.5, 11.6, 11.0, 10.8; LRMS (FAB+) 287; HRMS (FAB+) m/z: calcd for $C_{16}H_{31}O_4$ 287.2222 [M + H⁺], found 287.2216.

THP-Protected β -Hydroxy Ester **S5**. A screw cap test tube was charged with the core skeleton 35 (11 mg, 24 μ mol), THP-protected β -hydroxy carboxylic acid 54 (6.4 mg, 24 μ mol), and CH₂Cl₂ (120 μ L). To the solution was added DMAP (3.0 mg, 24 μ mol) and WSCD·HCl (9.3 mg, 49 μ mol) at 0 °C. After stirring at reflux for 2 h, the reaction was quenched with 1 M aqueous HCl. The resulting mixture was extracted with CH2Cl2 three times. The combined organic extracts were washed with H2O and brine, dried over Na2SO4, filtered, and concentrated under reduced pressure to give a residue, which was purified by preparative TLC (CH₂Cl₂-acetone = 40:1) to afford THPprotected β -hydroxy ester S5 (5.4 mg, 7.6 μ mol, 31%) as a pale yellow solid with recovery of the core skeleton 35 (3.5 mg, 7.8 μ mol, 32%). R_f = 0.45 (CH₂Cl₂-acetone =10:1); $[\alpha]_D^{27}$ -235 (c 0.215, CHCl₃); IR (neat, cm⁻¹): 3435, 2961, 2926, 2876, 1732, 1695, 1676, 1674, 1666, 1659, 1385, 1198, 1128, 1026, 756; 1 H NMR (600 MHz, CDCl₃): δ 6.58 (s, 1H), 6.52 (s, 1H), 6.33-6.27 (m, 1H), 6.23-6.19 (m, 1H), 5.96-5.93 (m, 0.4H), 5.92-5.87 (m, 0.6H), 5.14-5.09 (m, 1H), 4.93 (dd, J = 8.4, 1.8 Hz, 1H), 4.90 (d, J = 8.4 Hz, 1H), 4.71 (dd, J = 7.8,1.8 Hz, 1H), 4.67 (br s, 1H), 4.61 (ddd, J = 13.8, 7.2, 4.8 Hz, 1H), 4.45-4.39 (m, 1H), 3.96-3.87 (m, 0.6H), 3.87-3.80 (m, 0.4H), 3.50-3.43 (m, 0.6H), 3.41-3.35 (m, 0.4H), 3.13-2.94 (m, 4H), 2.71-2.66 (m, 1H), 2.29 (s, 3.6H), 2.28 (s, 2.4H), 1.83-1.74 (m, 1H), 1.72–1.65 (m, 1H), 1.60–1.38 (m, 8H), 1.30–1.16 (m, 1H), 1.22 (d, J = 7.2 Hz, 1.2 H), 1.15 (d, J = 7.2 Hz, 1.8 H), 1.08 - 1.00 (m, 1 H), 1.00 -0.82 (m, 10H); 13 C NMR (150 MHz, CDCl₃): δ 175.3, 174.2, 167.0, 166.9, 164.2, 164.1, 139.7, 139.3, 138.0, 137.90, 137.86, 137.6, 137.5, 110.9, 109.6, 109.4, 108.0, 107.9, 106.0, 105.6, 99.8, 99.0, 83.4, 81.2, 72.61, 72.56, 71.4, 71.0, 70.5, 69.4, 69.3, 64.1, 63.0, 62.5, 60.9, 60.6, 41.0, 40.55, 40.51, 40.48, 40.2, 40.10, 40.05, 34.8, 34.2, 31.9, 31.7, 30.9, 28.4, 28.0, 25.5, 25.4, 20.4, 20.1, 20.0, 19.7, 16.8, 16.5, 14.91, 14.86, 14.72, 14.70, 11.6, 11.1, 11.0, 10.6 (six signals are missing due to overlap); HRMS (ESI⁺) m/z: calcd for $C_{36}H_{50}N_2O_9S_2Na$ 741.2855 [M + Na⁺], found 741.2832.

β-Hydroxy Ester 51. A screw cap test tube was charged with THP-protected β-hydroxy ester S5 (2.7 mg, 3.9 μmol), MeOH (25 μL), and CH₂Cl₂ (25 μL). To the solution was added PPTS (2.0 mg, 7.8 μmol). After stirring at 50 °C for 1.5 h, the reaction mixture was diluted with CH₂Cl₂. The separated organic layer was washed twice with 13.2%

aqueous sodium chloride, dried over Na2SO4, filtered, and concentrated under reduced pressure to give a crude material, which was purified by preparative TLC (CH₂Cl₂-acetone = 20:1) to afford β hydroxy ester 51 (1.7 mg, 8.3 μ mol, 72%) as a white solid. R_i = 0.14 $(CH_2Cl_2$ -acetone = 20:1); $[\alpha]_D^{25}$ -330 (c 0.151, CHCl₃); IR (neat, cm⁻¹): 3468, 2961, 2926, 2874, 1732, 1693, 1680, 1676, 1666, 1651, 1393, 1385, 1339, 1192, 1126, 754; 1 H NMR (500 MHz, CDCl₃): δ 6.62-6.58 (m, 1H), 6.54-6.51 (m, 1H), 6.31 (dd, I = 8.5, 2.5 Hz, 1H), 6.22 (dd, J = 8.0, 2.5 Hz. 1H), 5.80 (ddd, J = 8.5, 2.0, 2.0 Hz, 1H), 5.22-5.17 (m, 1H), 4.93 (dd, J = 8.0, 2.5 Hz, 1H), 4.93-4.87(m, 1H), 4.72-4.64 (m, 1H), 4.66 (dd, J = 8.5, 2.0 Hz, 1H), 4.43 (br s, 1H), 4.72-4.64 (m, 1H), 4.66 (dd, J = 8.5, 2.0 Hz, 1H), 4.43 (br s, 1H), 4.72-4.64 (m, 1H), 4.66 (dd, J = 8.5, 2.0 Hz, 1H), 4.43 (br s, 1H), 4.66 (dd, J = 8.5, 2.0 Hz, 1H), 4.43 (br s, 1H), 4.66 (dd, J = 8.5, 2.0 Hz, 1H), 4.43 (br s, 1H), 4.66 (dd, J = 8.5, 2.0 Hz, 1H), 4.43 (br s, 1H), 4.66 (dd, J = 8.5, 2.0 Hz, 1H), 4.43 (br s, 1H), 4.66 (dd, J = 8.5, 2.0 Hz, 1H), 4.43 (br s, 1H), 4.66 (dd, J = 8.5, 2.0 Hz, 1H), 4.43 (br s, 1H), 4.66 (dd, J = 8.5, 2.0 Hz, 1H), 4.43 (br s, 1H), 4.66 (dd, J = 8.5, 2.0 Hz, 1H), 4.43 (br s, 1H), 4.66 (dd, J = 8.5, 2.0 Hz, 1H), 4.43 (br s, 1H), 4.66 (dd, J = 8.5, 2.0 Hz, 1H), 4.43 (br s, 1H)1 H), 3.67 (dd, J = 8.5, 2.5 Hz, 1H), 3.13–2.95 (m, 4H), 2.69 (dq, J = 7.5, 3.0 Hz, 1H), 2.53 (br s, 1H), 2.299 (s, 3H), 2.297 (s, 3H), 1.69 (ddd, J = 13.0, 9.0, 3.0 Hz, 1H), 1.65-1.39 (m, 3H), 1.20 (d, J = 7.0)Hz, 3H), 1.07-0.99 (m, 1H), 0.97-0.90 (m, 1H), 0.89 (d, J = 6.5 Hz, 3H), 0.86 (dd, J = 7.5, 7.5 Hz, 3H), 0.83 (d, J = 7.0 Hz, 3H); ¹³C NMR (150 MHz, CDCl₃): δ 175.5, 166.8, 164.4, 139.8, 138.05, 137.95, 137.6, 110.9, 109.5, 107.8, 105.7, 75.9, 72.6, 72.2, 70.4, 69.4, 64.1, 60.4, 42.0, 40.5, 40.4, 40.3, 33.6, 31.8, 27.9, 20.5, 16.1, 14.9, 14.8, 11.0, 9.6; HRMS (ESI⁺) m/z: calcd for C₃₁H₄₂N₂O₈S₂Na 657.2280 [M + Na⁺], found 657.2273.

 β -Keto Ester 37. A screw cap test tube was charged with the β hydroxy ester 43 (2.9 mg, 4.6 μ mol) and dry CH₂Cl₂ (76 μ L). To the solution was added Dess-Martin periodinane (2.3 mg, 9.1 μ mol) at 0 °C. After stirring at 0 °C for 1.5 h, the separated organic layer was washed with saturated aqueous NaHCO3 and brine, dried over Na₂SO₄, filtered, and concentrated under reduced pressure to give a crude material, which was purified by preparative TLC (CH₂Cl₂acetone = 20:1) to afford a 2:3 mixture of β -keto ester 37 and its C2' epimer, 38 (1.0 mg, 1.6 μ mol, 34%) as a pale yellow solid with recovery of the diol (1.2 mg, 1.9 μ mol, 41%). $R_f = 0.43$ (CH₂Cl₂acetone = 10:1). The physical data of a 2:3 mixture of 37 and 38: IR (neat, cm⁻¹): 3410, 2963, 2925, 2874, 2853, 1744, 1693, 1680, 1674, 1666, 1658, 1651, 1385, 1339, 1192, 1128, 1005, 754; ¹H NMR (600 MHz, CDCl₃): δ 6.62–6.58 (m, 1H), 6.53 (s, 1H), 6.33–6.27 (m, 1H), 6.21 (d, J = 7.8, 1H), 5.95-5.90 (m, 0.6H), 5.75-5.71 (m, 0.4H), 5.25 (d, J = 7.2 Hz, 0.4H), 5.11 (d, J = 8.4 Hz, 0.6H), 4.95-4.92 (m, 1H), 4.92-4.88 (m, 1H), 4.73-4.70 (m, 0.4H), 4.68 (dd, J = 9.0, 8.4Hz, 1H), 4.63-4.60 (m, 0.6H), 4.45 (m, 0.4H), 4.39 (m, 0.6H), 3.70 (q, J = 7.2 Hz, 0.6H), 3.65 (q, J = 7.2 Hz, 0.4H), 3.13-2.87 (m, 5H),2.32-2.26 (m, 6H), 1.80-1.70 (m, 1H), 1.41 (d, J = 6.6 Hz, 1.2H), 1.41-1.25 (m, 2H), 1.32 (d, J = 7.2 Hz, 1.8H), 1.17 (d, J = 7.2 Hz, 1.8H), 1.16-1.04 (m, 2H), 1.08 (d, J = 7.2 Hz, 1.2H), 0.91-0.84 (m, 6H); 13 C NMR (150 MHz, CDCl₃): δ 210.5, 210.0, 169.7, 169.4, 166.80, 166.78, 164.3, 164.2, 139.9, 139.6, 138.1, 138.0, 137.7, 110.9, 110.8, 109.4, 109.3, 107.9, 107.8, 105.4, 105.0, 73.0, 72.6, 72.5, 71.9, 70.5, 70.3, 69.6, 69.3, 64.1, 60.7, 60.3, 51.7, 50.9, 43.9, 43.1, 40.6, 40.3, 40.2, 40.0, 39.9, 32.2, 32.0, 29.4, 29.3, 19.5, 19.4, 17.4, 17.3, 14.94, 14.88, 14.78, 14.6, 12.7, 11.2, 11.1 (Six signals are missing due to overlap.); HRMS (ESI⁺) m/z: calcd for $C_{31}H_{40}N_2O_8S_2Na$ 655.2124 $[M + Na^{+}]$, found 655.2148.

β-Keto Ester 38. A screw cap test tube was charged with β-hydroxy ester 51 (1.5 mg, 2.4 μ mol) and dry CH₂Cl₂ (40 μ L). To the solution was added Dess–Martin periodinane (1.2 mg, 4.7 μ mol) at 0 °C. After stirring at 0 °C for 45 min, additional Dess-Martin periodinane (1.2 mg, 4.7 μ mol) was added at 0 °C, and the resulting mixture was stirred at 0 °C for 15 min. The reaction mixture was diluted with CH₂Cl₂. The organic layer was washed with saturated aqueous NaHCO3 and brine, dried over Na2SO4, filtered, and concentrated under reduced pressure to give a crude material, which was purified by preparative TLC (CH₂Cl₂-acetone = 20:1) to afford a 5:1 mixture of 38 and its C2' epimer, 37 (1.0 mg, 1.6 μ mol, 67%) as a white solid with recovery of β-hydroxy ester **51** (0.50 mg, 0.65 μ mol, 27%). R_f = 0.45 (CH₂Cl₂acetone =10:1); $[\alpha]_D^{27}$ -220 (c 0.090, CHCl₃); IR (neat, cm⁻¹): 3410, 2963, 2925, 2874, 2853, 1746, 1693, 1678, 1674, 1666, 1645, 1385, 1339, 1192, 1128, 1005, 752; ¹H NMR (600 MHz, CDCl₃): δ 6.61– 6.58 (m, 1H), 6.53 (s, 1H), 6.30 (dd, *J* = 7.8, 2.4 Hz, 1H), 6.22 (dd, *J* = 8.4, 2.4 Hz, 1H), 5.92 (ddd, J = 8.4, 2.4, 2.4 Hz, 1H), 5.11 (d, J = 9.0)Hz, 1H), 4.93 (dd, J = 8.4, 2.4 Hz, 1H), 4.92-4.88 (m, 1H), 4.67 (d, J= 7.8 Hz, 1H, 4.62 (dd, J = 7.8, 1.8 Hz, 1H, 4.39 (br s, 1H), 3.70 (q,) J = 7.2 Hz, 1H), 3.13–2.99 (m, 4H), 2.99–2.89 (m, 1H), 2.31 (s, 3H), 2.29 (s, 3H), 1.74 (ddd, J = 13.8, 7.8, 5.4 Hz, 1H), 1.43–1.24 (m, 2H), 1.32 (d, J = 7.2 Hz, 3H), 1.17 (d, J = 7.2 Hz, 3H), 1.16–1.04 (m, 2H), 0.91–0.80 (m, 6H); 13 C NMR (125 MHz, CDCl₃): δ 210.5, 169.3, 166.8, 164.3, 139.9, 138.1, 138.0, 137.7, 110.8, 109.3, 107.9, 105.0, 72.5, 71.9, 70.3, 69.6, 64.1, 60.7, 50.9, 43.8, 40.6, 40.2, 40.0, 32.2, 29.3, 19.3, 17.3, 14.9, 14.5, 12.8, 11.1; HRMS (ESI⁺) m/z: calcd for $C_{31}H_{40}N_2O_8S_2Na$ 655.2124 [M + Na⁺], found 655. 2105.

(+)-(2S,3R,4R,6R)-Oxazolidinone S6. A flame-dried 30 mL twonecked round-bottomed flask was charged with oxazolidinone 45 (400 mg,1.32 mmol), dry MeOH (266 μ L, 6.59 mmol), and dry THF (6.6 mL). To the solution was added LiBH₄ (143 mg, 6.59 mmol) at 0 $^{\circ}$ C. After stirring at 0 °C for 11.5 h, the reaction was quenched with 1 M NaOH aq. at 0 °C. The reaction mixture was diluted with water and extracted with EtOAc three times. The combined organic extracts were dried over Na2SO4, filtered, and concentrated under reduced pressure at 0 °C to give a crude material, which was purified by flash column chromatography (hexanes-EtOAc = 3:1) to afford alcohol 46 as a hexanes-EtOAc solution. All the solvent was not removed due to the volatility of the product. $R_f = 0.22$ (hexanes-EtOAc = 5:1). A flamedried 20 mL two-necked round-bottomed flask was charged with a hexanes-EtOAc solution of 46, Et₃N (918 μL, 6.59 mmol), dry DMSO (1.3 mL), and dry CH₂Cl₂ (2.6 mL). To the solution was added SO₃· Py (1.05 g, 6.59 mmol) at 0 °C. After stirring at 0 °C for 15 min, the reaction was quenched with water at 0 °C. The reaction mixture was extracted with CH2Cl2 three times. The combined organic extracts were dried over Na2SO4, filtered, and concentrated under reduced pressure at 0 °C to give a crude material, which was purified by flash column chromatography (hexanes-EtOAc = 3:1) to afford aldehyde 47 in hexanes-EtOAc solution. All the solvent was not removed due to the volatility of the product. $R_f = 0.42$ (hexanes-EtOAc = 5:1). A flamedried 20 mL two-necked round-bottomed flask was charged with (3S)-3-propionyl-4-benzyloxazolidin-2-one (338 mg, 1.45 mmol) and dry CH₂Cl₂ (1.3 mL). To the solution was added n-Bu₂BOTf (469 mg, 1.71 mmol) at 0 °C. After stirring at 0 °C for 10 min, Et₃N (239 μ L, 1.71 mmol) was added at 0 °C. Then, after stirring at 0 °C for 10 min, a CH₂Cl₂ solution of aldehyde 47 was added dropwise at −78 °C and stirring was continued at -78 °C for 1 h and at 0 °C for 0.5 h. The reaction was quenched with pH 7.0 phosphorus buffer (1.5 mL) and MeOH (4 mL), then MeOH-34% H₂O₂ aq. (2:1, 6 mL) at 0 °C. After stirring at 0 °C for 1 h, the reaction mixture was diluted with water and extracted with Et2O three times. The combined organic extracts were washed with 5% NaHCO3 brine, dried over Na2SO4, filtered, and concentrated under reduced pressure at 0 °C to give a crude material, which was purified by flash column chromatography (CH₂Cl₂ only) to afford the corresponding aldol product S6 (174 mg, 482 μ mol, 33% over 3 steps from 45) as a colorless solid. $R_f = 0.15$ (hexanes-EtOAc = 5:1); m.p.: 88.9–89.4 °C (hexanes, white prism); $[\alpha]_D^{27}$ +57.1 (c 1.26, CHCl₃). Spectral data matched the compound S4.

(+)-(2S,3R,4R,6R)-THP-Protected β -Hydroxy Carboxylic Acid **S7**. A flame-dried 20 mL two-necked round-bottomed flask was charged with the above aldol product S6 (174 mg, 482 mmol), DHP (132 μ L,1.45 mmol), and dry CH_2Cl_2 (2.5 mL). To the solution was added PPTS (48 mg, 0.19 mmol). After stirring at room temperature for 1.5 h, the reaction mixture was diluted with CH₂Cl₂. The separated organic layer was washed twice with 13.2% aqueous sodium chloride, dried over Na₂SO₄, filtered, and concentrated under reduced pressure to give a crude material as a colorless oil, which was used to the next reaction without further purification. $R_f = 0.33$ (hexanes-EtOAc = 5:1). A 20 mL two-necked round-bottomed flask was charged with the crude material, dry THF (1.8 mL), and H₂O (0.6 mL). To the solution was added H₂O₂ (34% w/w solution in H₂O, 0.24 mL, 2.4 mmol) and LiOH·H₂O (40 mg, 0.96 mmol) at 0 °C. After stirring at room temperature for 10.5 h, the reaction was quenched with saturated aqueous Na₂S₂O₃ and 1 M aqueous HCl. The aqueous layer was extracted with EtOAc three times. The combined organic extracts were dried over Na2SO4, filtered, and concentrated under reduced pressure to give a crude material, which was purified by flash column chromatography (hexanes-EtOAc = 3:1) to afford the THP-protected β -hydroxy carboxylic acid S7 (94.2 mg, 328 mmol, 68% over 2 steps

from S6) as a colorless oil. $R_f = 0.15$ (hexanes-EtOAc = 5:1); $[\alpha]_D^{27} + 21.0$ (c 4.40, CHCl₃). Spectral data matched the compound 54.

THP-Protected β-Hydroxy Ester **S8**. A screw cap test tube was charged with the core skeleton 35 (11 mg, 24 μ mol), the above THPprotected β -hydroxy carboxylic acid S7 (6.4 mg, 24 μ mol), and CH_2Cl_2 (120 μ L). The solution was cooled to 0 °C. To the solution was added DMAP (3.0 mg, 24 μ mol) and WSCD·HCl (9.3 mg, 49 μ mol) at 0 °C. After stirring the resulting mixture at reflux for 3 h, the reaction was quenched with 1 M aqueous HCl. The resulting mixture was extracted with CH2Cl2 three times. The combined organic extracts were washed with H2O and brine, dried over Na2SO4, filtered, and concentrated under reduced pressure to give a residue, which was purified by preparative TLC (CH₂Cl₂-EtOAc = 40:1) to afford THPprotected β -hydroxy ester S8 (6.4 mg, 8.9 μ mol, 37%) as a colorless oil with recovery of the core skeleton 35 (3.5 mg, 7.8 μ mol, 32%). $R_f =$ 0.47 (CH₂Cl₂-acetone = 10:1); $[\alpha]_D^{27}$ -196 (c 0.300, CHCl₃); IR (neat, cm⁻¹): 3420, 2961, 2926, 2876, 1732, 1691, 1676, 1668, 1662, 1654, 1385, 1198, 1126, 1026, 756; 1 H NMR (500 MHz, CDCl₃): δ 6.60-6.56 (m, 1H), 6.52 (s, 1H), 6.32-6.25 (m, 1H), 6.24-6.19 (m, 1H), 5.83 (ddd, *J* = 7.5, 2.0, 2.0 Hz, 0.4H), 5.76 (ddd, *J* = 8.0, 2.0, 2.0 Hz, 0.6H), 5.23-5.08 (m, 1H), 4.95-4.86 (m, 1.6H), 4.84 (dd, J =8.0, 2.0 Hz, 0.4H), 4.70-4.59 (m, 2H), 4.52-4.40 (m, 1H), 3.97-3.82 (m, 2H), 3.51–3.43 (m, 0.6H), 3.41–3.35 (m, 0.4H), 3.12–2.95 (m, 4H), 2.69-2.59 (m, 1H), 2.30-2.26 (m, 6H), 1.85-1.75 (m, 1H), 1.75-1.65 (m, 1H), 1.60-1.35 (m, 8H), 1.28 (d, J = 7.0 Hz, 1.2H), 1.25-1.15 (m, 1H), 1.24 (d, J = 7.0 Hz, 1.8H), 1.08-0.78 (m, 11H); 13 C NMR (125 MHz, CDCl₃): δ 174.8, 174.1, 166.94, 166.87, 164.2, 164.1, 139.6, 139.2, 137.93, 137.86, 137.6, 110.84, 110.81, 109.6, 109.4, 107.9, 107.8, 106.2, 105.7, 101.6, 100.1, 84.1, 81.8, 72.6, 72.1, $71.7,\ 70.5,\ 70.4,\ 69.4,\ 64.1,\ 63.9,\ 63.2,\ 60.7,\ 60.4,\ 41.73,\ 41.70,\ 40.5,$ 40.43, 40.35, 40.32, 40.0, 34.9, 33.8, 31.79, 31.76, 31.3, 31.1, 28.3, 28.0, 25.5, 25.3, 20.9, 20.4, 20.19, 20.16, 16.60, 16.58, 14.86, 14.81, 14.7, 11.7, 11.1, 10.9, 10.3, 9.0 (Seven signals are missing due to overlap.); HRMS (ESI⁺) m/z: calcd for $C_{36}H_{50}N_2O_9S_2K$ 757.2595 [M + K⁺], found 757.2612.

β-Hydroxy Ester 52. A screw cap test tube was charged with the above THP-protected β -hydroxy ester S8 (6.0 mg, 8.4 μ mol), MeOH (50 μ L), and CH₂Cl₂ (50 μ L). To the solution was added PPTS (4.2 mg, 17 μ mol). After stirring at 50 °C for 2.5 h, the reaction mixture was diluted with CH2Cl2. The separated organic layer was washed twice with 13.2% aqueous sodium chloride, dried over Na2SO4, filtered, and concentrated under reduced pressure to give a crude material, which was purified by preparative TLC (CH₂Cl₂-acetone = 20:1) to afford β -hydroxy ester 52 (3.0 mg, 4.7 μ mol, 56%) as a white solid. $R_f = 0.39$ (CH₂Cl₂-acetone = 10:1); $[\alpha]_D^{25}$ -277 (c 0.125, CHCl₃); IR (neat, cm⁻¹): 3501, 2961, 2924, 2874, 1728, 1691, 1666, 1651, 1393, 1385, 1194, 1126, 978, 756; ¹H NMR (600 MHz, CDCl₃): δ 6.61 (s, 1H), 6.51 (s, 1H), 6.33 (dd, J = 9.4, 2.4 Hz, 1H), 6.21 (dd, *J* = 8.4, 2.4 Hz, 1H), 5.89 (ddd, *J* = 7.8, 1.8, 1.8 Hz, 1H), 5.15 (d, J = 8.4 Hz, 1H), 4.93 (dd, J = 8.4, 2.4 Hz, 1H), 4.89 (d, J = 7.2 Hz, 1Hz)1H), 4.68 (d, J = 7.2 Hz, 1H), 4.62 (dd, J = 8.4, 2.4 Hz, 1H), 4.43 (br s, 1H), 3.77 (dd, *J* = 9.0, 1.2 Hz, 1H), 3.12–2.96 (m, 5H), 2.69 (dq, *J* = 7.2, 1.2 Hz, 1H), 2.32 (s, 3H), 2.31 (s, 3H), 1.76 (ddd, J = 13.8, 9.6,3.6 Hz, 1H), 1.62–1.41 (m, 3H), 1.21 (d, J = 6.6 Hz, 3H), 1.03 (dq, J= 13.2, 7.2 Hz, 1H), 0.92 (ddd, J = 13.8, 9.6, 4.2 Hz, 1H), 0.89 (d, J = 7.2 Hz, 3H), 0.86 (d, J = 7.2 Hz, 3H), 0.80 (d, J = 6.0 Hz, 3H); ¹³C NMR (150 MHz, CDCl₃): δ 175.4, 166.8, 164.9, 139.9, 138.2, 137.6, 110.8, 109.3, 107.8, 105.6, 75.0, 72.5, 71.8, 70.2, 69.6, 64.2, 60.8, 42.1, 41.0, 40.7, 40.3, 33.2, 31.8, 28.0, 20.5, 15.9, 14.9, 14.8, 11.1, 9.0: HRMS (ESI⁺) m/z: calcd for $C_{31}H_{42}N_2O_8S_2Na$ 657.2280 [M + Na⁺], found 657.2252.

β-Keto Ester **39**. A screw cap test tube was charged with β-hydroxy ester **52** (2.5 mg, 3.9 μmol) and dry CH₂Cl₂ (40 μL). To the solution was added Dess—Martin periodinane (2.0 mg, 7.9 μmol) at 0 °C. After stirring at 0 °C for 1 h, additional Dess—Martin periodinane (1.0 mg, 3.9 μmol) was added, and the resulting mixture was stirred at 0 °C for 1 h. The reaction mixture was diluted with CH₂Cl₂. The separated organic layer was washed with saturated aqueous NaHCO₃ and brine, dried over Na₂SO₄, filtered, and concentrated under reduced pressure to give a crude material, which was purified by preparative TLC

(CH₂Cl₂-acetone = 20:1) to afford a 3:2 mixture of 39 and its C2' epimer 40 (1.2 mg, 1.9 μ mol, 48%) as a pale yellow solid with recovery of β -hydroxy ester **52** (0.90 mg, 1.4 μ mol, 36%). $R_f = 0.45$ (CH₂Cl₂acetone = 10:1). The physical data of 3:2 mixture of 39 and 40: IR (neat, cm⁻¹): 3400, 2963, 2924, 2874, 2853, 1738, 1712, 1693, 1680, 1674, 1666, 1658, 1645, 1385, 1339, 1194, 1126, 1003, 752; ¹H NMR (600 MHz, CDCl₃): δ 6.62–6.58 (m, 1H), 6.53 (s, 1H), 6.32 (dd, J =8.4, 2.4 Hz, 0.6H), 6.29 (dd, I = 8.4, 2.4 Hz, 0.4H), 6.22 (dd, I = 8.4, 2.4 Hz, 1H), 5.92 (ddd, J = 8.4, 2.4, 2.4 Hz, 0.4H), 5.74 (ddd, J = 7.8, 2.4, 2.4 Hz, 0.6H), 5.27-5.22 (m, 0.6H), 5.12-5.08 (m, 0.4H), 4.95-4.92 (m, 1H), 4.92-4.88 (m, 1H), 4.72 (dd, J = 8.4, 1.8 Hz, 0.6H), 4.71-4.65 (m, 1H), 4.60 (dd, J = 8.4, 1.8 Hz, 0.4H), 4.49-4.44 (m, 0.6H), 4.42-4.37 (m, 0.4H), 3.72 (q, J = 7.2 Hz, 0.4H), 3.64 (q, J =7.2 Hz, 0.6H), 3.13-2.98 (m, 4.4H), 2.90 (dq, J = 13.8, 6.6 Hz, 0.6H), 2.33-2.31 (m, 3H), 2.30 (s, 1.8H), 2.29 (s, 1.2H), 1.82 (ddd, J = 13.8, 8.4, 5.4 Hz, 0.4H), 1.67 (ddd, *J* = 13.2, 7.8, 6.0 Hz, 0.6H), 1.40 (d, *J* = 7.2 Hz, 1.8H), 1.40–1.24 (m, 3H), 1.33 (d, J = 7.2 Hz, 1.2H), 1.13 (d, J = 7.2 Hz, 1.8H), 1.12 (d, J = 7.2 Hz, 1.2H), 1.11–1.03 (m, 1H), 0.92-0.84 (m, 6H); ¹³C NMR (150 MHz, CDCl₃): δ 210.20, 210.17, 169.6, 169.3, 166.82, 166.79, 164.3, 164.2, 139.9, 139.6, 138.1, 138.0, 137.7, 110.8, 109.4, 109.3, 107.9, 107.8, 105.3, 105.0, 73.0, 72.62, 72.57, 71.9, 70.44, 70.38, 69.5, 69.4, 64.2, 64.1, 60.7, 60.4, 52.0, 50.4, 44.0, 43.3, 40.5, 40.4, 40.3, 40.2, 39.7, 32.1, 29.7, 29.5, 20.2, 19.52, 19.49, 17.3, 17.0, 14.91, 14.88, 14.77, 14.6, 13.0, 12.9, 11.2, 11.1 (five signals are missing due to overlap); HRMS (ESI⁺) m/z: calcd for $C_{31}H_{40}N_2O_8S_2Na$ 655.2124 [M + Na⁺], found 655.2120.

THP-Protected β -Hydroxy Ester S9. A screw cap test tube was charged with 35 (14 mg, 31 μ mol), 50 (8.9 mg, 31 μ mol), and CH₂Cl₂ (80 μ L). To the solution was added DMAP (3.8 mg, 31 μ mol) and WSCD·HCl (11.9 mg, 62.1 μ mol) at 0 °C. After stirring at reflux for 2 h, the reaction was quenched with 1 M aqueous HCl. The resulting mixture was extracted with CH2Cl2 three times. The combined organic extracts were washed with H₂O and brine, dried over Na₂SO₄, filtered, and concentrated under reduced pressure. The residue was purified by preparative TLC (CH₂Cl₂-acetone = 20:1) to afford the desired THPprotected β -hydroxy ester **S9** (8.0 mg, 11 μ mol, 36%) as a pale yellow solid with recovery of 35 (7.0 mg, 16 μ mol, 50%). $R_f = 0.41$ (CH₂Cl₂acetone = 10:1); $[\alpha]_D^{27}$ -229 (c, 0.515, CHCl₃); IR (neat, cm⁻¹): 3400, 2957, 2928, 2874, 1732, 1693, 1681, 1666, 1658, 1651, 1385, 1198, 1128, 1026, 754, 667; ¹H NMR (500 MHz, CDCl₃): δ 6.61–6.57 (m, 1H), 6.54-6.49 (m, 1H), 6.33-6.25 (m, 1H), 6.22-6.18 (m, 1H), 5.90 (ddd, J = 8.0, 2.5, 2.5 Hz, 0.4H), 5.88 (ddd, J = 8.0, 2.5, 2.5 Hz, 0.6H), 5.23-5.10 (m, 1H), 4.93 (dd, J = 8.0, 2.0 Hz, 0.6H), 4.92-4.85 (m, 1H), 4.83 (dd, J = 8.0, 2.0 Hz, 0.4H), 4.70–4.63 (m, 1.2H), 4.62– 4.57 (m, 0.8H), 4.42 (br s, 1H), 3.95-3.84 (m, 1H), 3.84-3.80 (m, 1H), 3.48-3.42 (m, 0.5H), 3.42-3.36 (m, 0.5H), 3.14-2.95 (m, 4H), 2.75-2.65 (m, 1H), 2.30-2.26 (m, 6H), 1.85-1.75 (m, 1H), 1.75-1.65 (m, 3H), 1.65-1.42 (m, 6H), 1.42-1.33 (m, 1H), 1.23 (d, J = 7.0 (d, J = 7.0 (d, J = 7.0 (d, J = 7.0 (d, J = 7.0Hz, 1.2H), 1.17 (d, J = 7.0 Hz, 1.8H), 1.10–0.93 (m, 2H), 0.96 (d, J =7.0 Hz, 1.8H), 0.94 (d, J = 7.0 Hz, 1.2H), 0.90–0.78 (m, 7H); 13 C NMR (125 MHz, CDCl₃): δ 175.0, 174.4, 167.0, 166.9, 164.10, 164.07, 139.8, 139.4, 137.95, 137.86, 137.82, 137.61, 137.57, 110.8, 109.6, 109.5, 108.0, 107.9, 105.9, 105.6, 100.5, 99.4, 83.1, 81.1, 72.6, 71.5, 71.2, 70.51, 70.48, 64.12, 64.10, 63.2, 63.0, 60.7, 60.5, 41.8, 41.4, 41.1, 41.0, 40.6, 40., 40.4, 40.2, 34.4, 34.0, 31.7, 31.5, 30.95, 30.93, 28.9, 28.4, 25.44, 25.40, 20.2, 20.1, 20.0, 19.6, 15.8, 15.7, 14.86, 14.83, 14.7, 12.4, 11.8, 11.1, 11.0 (six signals are missing due to overlap); HRMS (ESI⁺) m/z: calcd for C₃₆H₅₀N₂O₉S₂Na 741.2855 [M + Na⁺], found 741.2835.

β-Hydroxy Ester **53**. A screw cap test tube was charged with the THP-protected β-hydroxy ester **S9** (4.0 mg, 5.6 μmol), MeOH (28 μL), and CH₂Cl₂ (28 μL). To the solution was added PPTS (2.8 mg, 11 μmol). After stirring at 50 °C for 1.5 h, the reaction mixture was diluted with CH₂Cl₂. The separated organic layer was washed twice with 13.2% aqueous sodium chloride, dried over Na₂SO₄, filtered, and concentrated under reduced pressure to give a crude material, which was purified by preparative TLC (CH₂Cl₂-acetone = 20:1) to afford β-hydroxy ester **53** (2.7 mg, 4.3 μmol, 76%) as a pale yellow solid. R_f = 0.34 (CH₂Cl₂-acetone = 10:1); $[\alpha]_D^{2D}$ -253 (c 0.530, CHCl₃); IR (neat, cm⁻¹): 3468, 2962, 2924, 2876, 1732, 1693, 1680, 1666, 1651,

1393, 1384, 1337, 1192, 1128, 756; 1 H NMR (500 MHz, CDCl₃): δ 6.61–6.57 (m, 1H), 6.54–6.51 (m, 1H), 6.31 (dd, J = 8.5, 2.5 Hz, 1H), 6.22 (dd, J = 8.0, 2.5 Hz, 1H), 5.79 (ddd, J = 8.0, 2.0, 2.0 Hz, 1H), 5.19 (d, J = 8.5 Hz, 1H), 4.93 (dd, J = 8.5, 2.0 Hz, 1H), 4.89 (d, J = 7.5 Hz, 1H), 4.71–4.66 (m, 1H), 4.64 (dd, J = 8.0, 2.0 Hz, 1H), 4.42 (d, J = 6.0 Hz, 1H), 3.67 (dd, J = 7.0, 6.0 Hz, 1H), 3.14–2.95 (m, 4H), 2.69 (ddd, J = 14.0, 7.5, 4.5 Hz, 1H), 2.30 (s, 3H), 2.29 (s, 3H), 1.68–1.59 (m, 1H), 1.50–1.37 (m, 2H), 1.27–1.19 (m, 1H), 1.22 (d, J = 7.0 Hz, 3H), 1.06–0.92 (m, 2H), 0.98 (d, J = 6.5 Hz, 3H), 0.88–0.83 (m, 6H); 13 C NMR (125 MHz, CDCl₃): δ 175.3, 166.8 164.3, 139.8, 138.00, 137.95, 137.6, 110.9, 109.6, 107.8, 105.6, 75.5, 72.6, 72.1, 70.4, 69.4, 64.1, 60.4, 42.3, 40.6, 40.4, 40.2, 33.3, 31.3, 28.1, 20.0, 15.1, 14.84, 14.80, 11.0, 10.9; HRMS (ESI⁺) m/z: calcd for $C_{31}H_{42}N_{2}O_{8}S_{2}Na$ 657.2280 [M + Na⁺], found 657.2268.

 β -Keto Ester 40. A screw cap test tube was charged with the β hydroxy ester 53 (2.7 mg, 4.3 μ mol) and dry CH₂Cl₂ (70 μ L). To the solution was added Dess-Martin periodinane (2.1 mg, 8.5 µmol) at 0 °C. After stirring at 0 °C for 1.5 h, the reaction mixture was diluted with CH2Cl2. The separated organic layer was washed with saturated aqueous NaHCO3 and brine, dried over Na2SO4, filtered, and concentrated under reduced pressure to give a crude material, which was purified by preparative TLC (CH₂Cl₂-acetone = 10:1) to afford a 5:1 mixture of β -keto ester 40 and its C2' epimer, 39 (1.0 mg, 1.6 μ mol, 37%) as a pale yellow solid with recovery of the β -hydroxy ester 53 (1.2 mg, 1.9 μ mol, 44%). The physical data of a 5:1 mixture of 39 and 40: $R_f = 0.43$ (CH₂Cl₂-acetone = 10:1); $[\alpha]_D^{27}$ -246 (c 0.210, CHCl₃); IR (neat, cm⁻¹): 3400, 2963, 2926, 2876, 1747, 1713, 1693, 1682, 1666, 1651, 1634, 1396, 1385, 1194, 1124, 1003, 754; ¹H NMR (600 MHz, CDCl₃): δ 6.60 (s, 1H), 6.53 (s, 1H), 6.29 (dd, J = 8.4, 2.4 Hz, 1H), 6.22 (dd, J = 8.4, 2.4 Hz, 1H), 5.96-5.92 (m, 1H), 5.14-5.09 (m, 1H,), 4.94 (dd, J = 8.4, 1.2 Hz, 1H), 4.93-4.87 (d, J = 7.8 Hz,1H), 4.71-4.65 (m, 1H), 4.61 (dd, J = 8.4, 1.8 Hz, 1H), 4.39 (br s, 1H), 3.72 (q, J = 7.2 Hz, 1H), 3.14-2.98 (m, 5H), 2.31 (s, 3H), 2.29(s, 3H), 1.87-1.79 (m, 1H), 1.43-1.25 (m, 2H), 1.33 (d, J = 7.2 Hz, 3H), 1.19-1.03 (m, 5H), 0.96-0.84 (m, 6H); ¹³C NMR (150 MHz, CDCl₃): δ 210.2, 169.3, 166.8, 164.3, 139.9, 138.1, 138.0, 137.6, 110.8, 109.4, 107.9, 105.0, 72.6, 71.9, 70.4, 69.5, 64.1, 60.7, 50.4, 44.0, 40.5, 40.2, 39.7, 32.1, 29.5, 19.5, 17.3, 14.9, 14.6, 13.0, 11.2; HRMS (ESI⁺) m/z: calcd for $C_{31}H_{40}N_2O_8S_2Na$ 655.2124 [M + Na⁺], found 655.2123.

THP-Protected β -Hydroxy Ester **55**. A screw cap test tube was charged with (-)-deacetylaranotin (5) (4.3 mg, 10 μ mol), THPprotected β -hydroxy carboxylic acid 54 (2.3 mg, 8.2 μ mol), and CH₂Cl₂ (200 µL). To the solution was added DMAP (1.2 mg, 10 μ mol) and WSCD·HCl (3.9 mg, 20 μ mol) at 0 °C. After stirring at reflux for 2.5 h, the reaction was quenched with 1 M aqueous HCl. The resulting mixture was extracted with CH₂Cl₂ three times. The combined organic extracts were washed with water and brine, dried over Na₂SO₄, filtered, and concentrated under reduced pressure. The residue was purified by preparative TLC (CH₂Cl₂-acetone = 10:1) to afford THP-protected β -hydroxy ester 55 (1.5 mg, 2.1 μ mol, 21%, d.r. = 5:1) as a white solid with recovery of (-)-deacetylaranotin (5) (1.9) mg, 4.4 mmol, 44%). $R_f = 0.59$ (CH₂Cl₂-acetone = 10:1); $[\alpha]_D^{27}$ -460 (c 0.044, CHCl₃); IR (neat, cm⁻¹): 3360, 2953, 2930, 2874, 2853, 1732, 1715, 1693, 1456, 1441, 1362, 1281, 1200, 1140, 1078, 1026, 997, 752. The ¹H and ¹³C NMR data of major diastereomer: ¹H NMR (500 MHz, CDCl₃): δ 6.66–6.62 (m, 1H), 6.58–6.53 (m, 1H), 6.31 (dd, J = 8.0, 2.0 Hz, 1H), 6.26 (dd, J = 8.5, 2.0 Hz, 1H), 5.92-5.89 (m,1H), 5.71 (dt, J = 9.0, 2.0 Hz, 1H), 5.10–5.05 (m, 1H), 4.90–4.86 (m, 0.5H), 4.79 (dd, J = 8.5, 2.0 Hz, 1H), 4.78-4.73 (m, 1.5H), 4.62-4.58(m, 2H), 4.08-4.04 (m, 1H), 3.95-3.81 (m, 2H), 3.47-3.43 (m, 1H), 3.08 (d, J = 18.5 Hz, 1H), 2.74 (d, J = 17.5 Hz, 1H), 2.68-2.62 (m, 1Hz, 11H), 1.82-1.75 (m, 2H), 1.72-1.63 (m, 1H), 1.60-1.38 (m, 6H), 1.20-1.12 (m, 1H), 1.11 (d, J = 7.0 Hz, 3H), 1.07-0.95 (m, 2H), 0.95(d, J = 7.0 Hz, 3H), 0.92–0.84 (m, 6H); ¹³C NMR (125 MHz, $CDCl_3$): δ 175.1, 165.4, 141.2, 139.5, 138.80, 138.76, 112.7, 109.7, 108.2, 105.2, 99.1, 81.4, 74.7, 70.7, 69.6, 67.9, 62.92, 62.88, 40.9, 39.9, 35.7, 34.8, 34.6, 31.9, 30.9, 29.7, 28.4, 25.5, 20.5, 20.2, 19.9, 16.7, 111.7, 11.1; HRMS (ESI⁺) m/z: calcd for C₃₄H₄₄N₂O₉S₂Na 711.2386 $[M + Na^{+}]$, found 711.2377.

β-Hydroxy Ester 56. A screw cap test tube was charged with THPprotected β -hydroxy ester 55 (2.7 mg, 3.9 μ mol), MeOH (25 μ L), and CH_2Cl_2 (25 μ L). To the solution was added PPTS (2.0 mg, 7.8 μ mol) at room temperature. After stirring at 50 °C for 1.5 h, the reaction mixture was diluted with CH₂Cl₂. The organic layer was washed twice with 13.2% aqueous sodium chloride, dried over Na2SO4, filtered, and concentrated under reduced pressure to give a crude material, which was used to the next reaction without further purification. The physical data of the purified β -hydroxy ester **56**: $R_f = 0.41$ (CH₂Cl₂-acetone = 10:1); $[\alpha]_D^{27}$ -380 (c 0.065, CHCl₃); IR (neat, cm⁻¹): 3508, 3350, 2959, 2928, 2874, 2853, 1715, 1693, 1682, 1454, 1371, 1281, 1184, 1140, 756; ¹H NMR (600 MHz, CDCl₃): δ 6.65 (d, J = 1.8 Hz, 1H), 6.57 (s, 1H), 6.32 (dd, J = 8.4, 3.0 Hz, 1H), 6.26 (dd, J = 8.4, 2.4 Hz, 1H), 5.87 (d, J = 1.8 Hz, 1H), 5.68 (dt, J = 8.4, 1.8 Hz, 1H), 5.16-5.12(m, 1H), 4.81–4.78 (m, 1H), 4.78–4.75 (m, 1H), 4.63–4.59 (m, 1H), 4.59-4.55 (m, 1H), 4.10-4.04 (m, 1H), 3.93 (dt, J = 17.4, 1.8 Hz, 1H), 3.66-3.63 (m, 1H), 3.09 (d, J = 18.6 Hz, 1H), 2.77 (d, J = 17.4Hz, 1H), 2.66 (ddd, J = 14.4, 7.8, 3.0 Hz, 1H), 2.37 (d, J = 4.8 Hz, 1H), 1.67-1.42 (m, 3H), 1.23-1.20 (m, 1H), 1.15 (d, J = 9.6 Hz, 3H), 1.08-0.99 (m, 1H), 0.95 (ddd, J = 13.2, 9.0, 4.2 Hz, 1H), 0.90 (d, I = 6.6 Hz, 3H), 0.87 (dd, I = 7.8, 6.6 Hz, 3H), 0.84 (d, I = 6.6 Hz, 3H)3H); 13 C NMR (150 MHz, CDCl₃): δ 175.2, 165.3, 144.4, 141.5, 139.7, 139.0, 117.6, 109.7, 105.2, 96.7, 93.8, 75.9, 74.8, 70.6, 70.2, 67.9, 62.6, 42.0, 41.2, 40.4, 35.7, 34.6, 33.8, 31.8, 28.0, 20.5, 16.2, 11.1, 9.5; HRMS (ESI⁺) m/z: calcd for $C_{29}H_{36}N_2O_8S_2N_a$ 627.1811 [M + Na⁺], found 627.1807.

Hirsutellomycin (57). A screw cap test tube was charged with the crude β -hydroxy ester **56** and dry CH_2Cl_2 (50 μ L). To the solution was added Dess-Martin periodinane (1.4 mg, 5.6 μ mol) at 0 °C. After stirring at 0 °C for 1.5 h, the reaction mixture was diluted with CH2Cl2. The organic layer was washed with saturated aqueous NaHCO3 and brine, dried over Na2SO4, filtered, and concentrated under reduced pressure to give a crude material, which was purified by preparative TLC (CH₂Cl₂-acetone = 20:1) to afford 57 (0.80 mg, 1.3 μ mol) as a colorless oil with recovery of β -hydroxy ester **56** (0.80 mg, 1.3 μ mol), which was placed in a flame-dried screw cap test tube and dissolved in dry CH₂Cl₂ (50 µL). To the solution was added Dess-Martin periodinane (0.70 mg, 2.6 μ mol) at 0 °C. After stirring at 0 °C for 1.5 h, the reaction mixture was diluted with CH2Cl2. The organic layer was washed with saturated aqueous NaHCO3, followed by brine, dried over Na2SO4, filtered, and concentrated under reduced pressure to give a crude material, which was purified by preparative TLC $(CH_2Cl_2$ -acetone = 20:1) to afford 57 (0.56 mg, 0.91 μ mol) as a pale yellow solid with recovery of 56 (0.18 mg, 0.030 μ mol). The combined yield of hirsutellomycin (57) was 1.26 mg (58%, over 3 steps from 55). Synthetic hirsutellomycin (57) existed as a 10:1 mixture of 57 and its C2' epimer 57. $R_f = 0.57$ (CH₂Cl₂-acetone = 10:1); $[\alpha]_D^{27}$ -410 (c 0.095, CHCl₃); IR (neat, cm⁻¹): 3342, 2961, 2930, 2874, 2851, 1747, 1715, 1693, 1682, 1454, 1371, 1188, 1140, 754; ¹H NMR (500 MHz, CDCl₃): δ 6.64 (d, J = 2.0 Hz, 1H), 6.58 (s, 1H), 6.31 (dd, J = 8.5, 2.5 Hz, 1H), 6.26 (dd, J = 8.5, 2.5 Hz, 1H), 5.92-5.85 (m, 1H), 5.78-5.74 (m, 1H), 5.07 (dd, J = 8.5, 1.5 Hz, 1H), 4.79 (dd, J = 8.5, 2.0 Hz, 1H), 4.79-4.74 (m, 1H), 4.64-4.58(m, 1H), 4.55 (dd, J = 8.5, 2.0 Hz, 1H), 4.05 (d, J = 17.5 Hz, 1H), 3.92(d, J = 17.0 Hz, 1H), 3.66 (q, J = 7.0 Hz, 1H), 3.08 (d, J = 18.5 Hz,1H), 2.92 (q, J = 7.0 Hz, 1H), 2.79 (d, J = 17.0 Hz, 1H,), 1.73 (ddd, J = 13.5, 7.5, 6.0 Hz, 1H), 1.42–1.15 (m, 2H), 1.28 (d, J = 7.0 Hz, 3H), 1.20-1.03 (m, 2H), 1.15 (d, J = 6.5 Hz, 3H), 0.89 (d, J = 6.5 Hz, 3H), 0.87 (t, J = 7.5 Hz, 3H); ¹³C NMR (150 MHz, CDCl₃): δ 210.4, 169.2, 165.3, 162.5, 141.4, 139.6, 138.8, 112.6, 109.7, 108.1, 104.8, 76.4, 74.8, 70.7, 69.9, 67.9, 62.9, 51.0, 43.7, 40.0, 35.8, 34.6, 32.2, 29.3, 19.4, 17.2, 12.8, 11.1 (one signal is missing due to overlap); HRMS (ESI⁺) m/z: calcd for $C_{29}H_{34}N_2O_8S_2Na$ 625.1654 [M + Na⁺], found 625.1661.

ASSOCIATED CONTENT

S Supporting Information

The Supporting Information is available free of charge on the ACS Publications website at DOI: 10.1021/acs.joc.6b02452.

Copies of ¹H and ¹³C NMR spectra. (PDF)

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Notes

The authors declare no competing financial interest.

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